

DNA damage and repair in cells exposed to radiation

Beata Brzozowska

Jerzy Pniewski and Leopold Infeld Colloquium, 4th of March, 2024



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Biomedical Physics Division \subset Faculty of Physics \subset University of Warsaw.

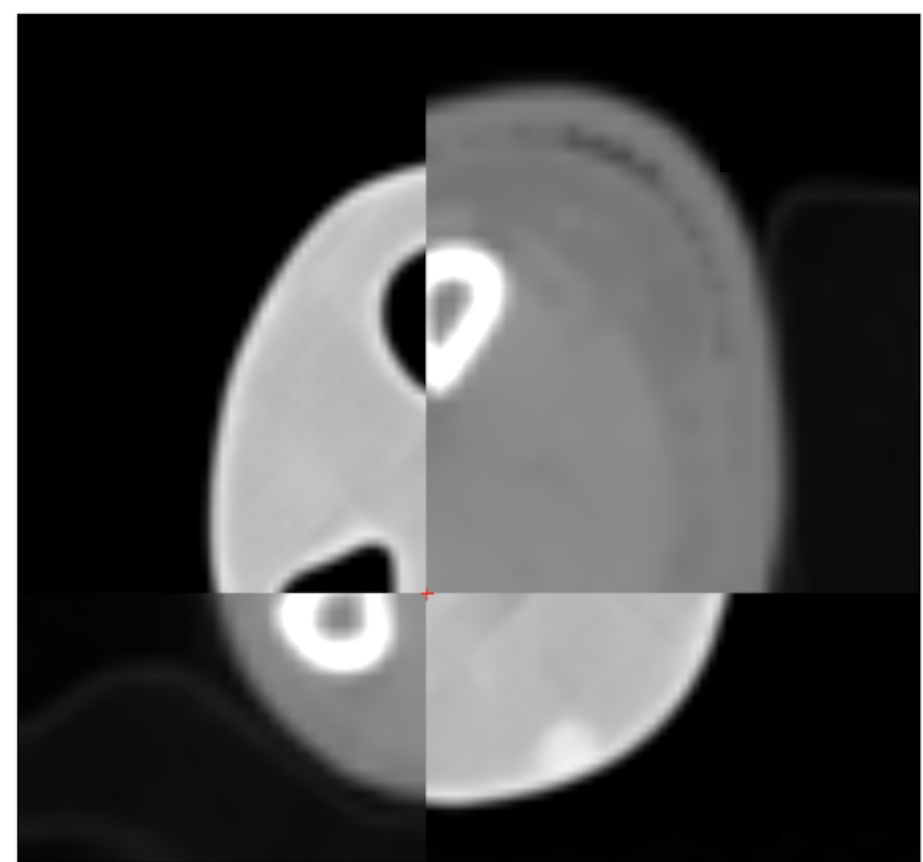
Neuroinformatics

Medical physics

Medical physics @ Biomedical Physics Division

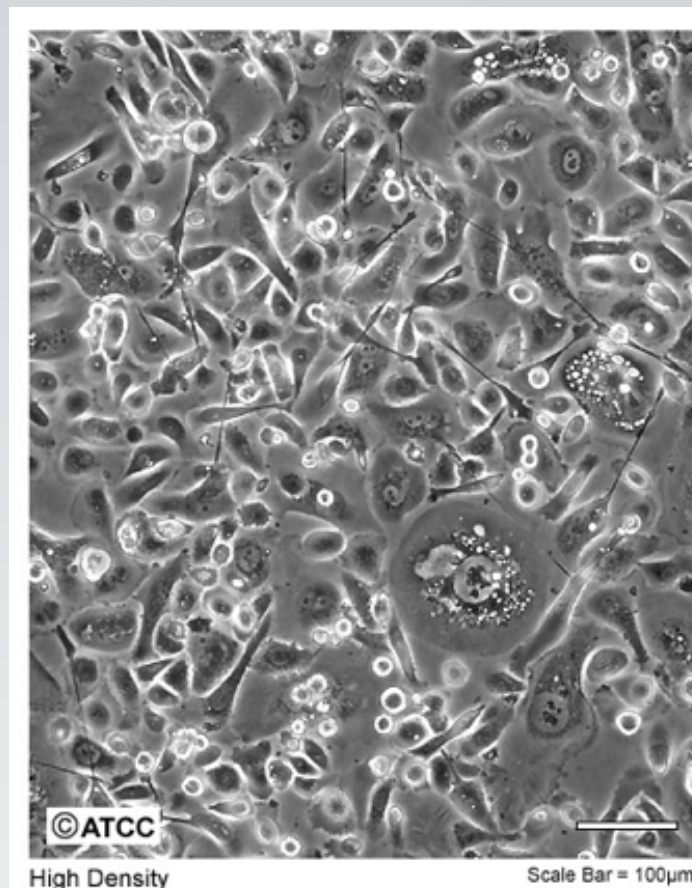
DNA damage and repair processes in cells exposed to ionising radiation

5-150 cm



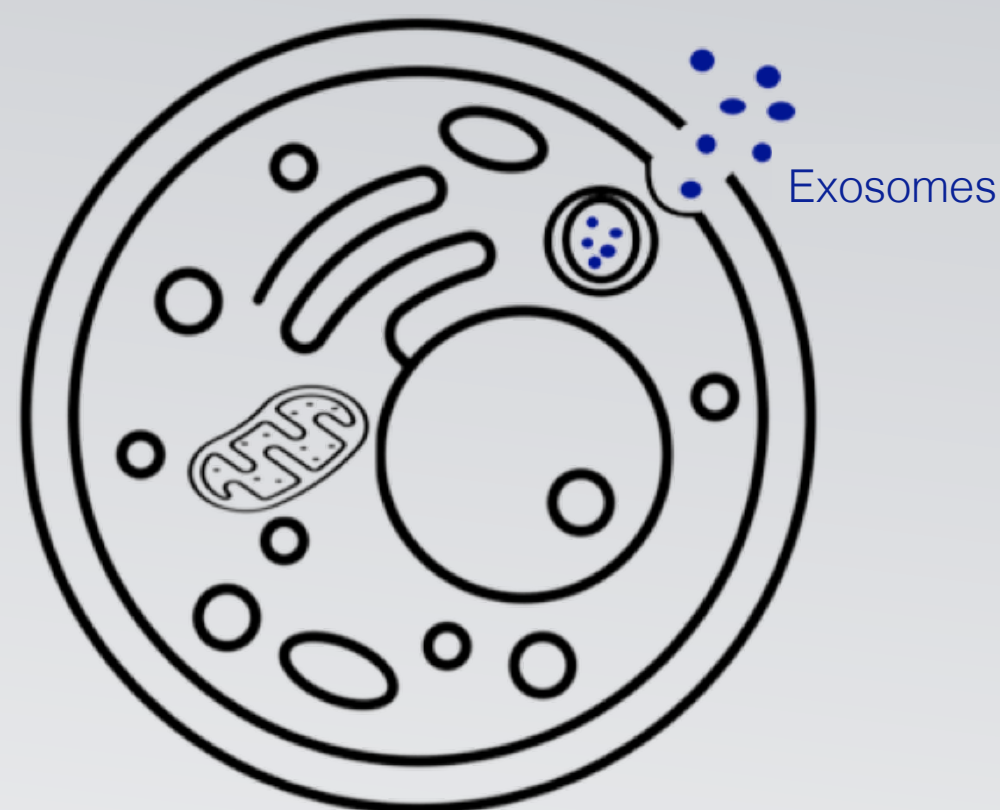
radiotherapy

5-50 μm

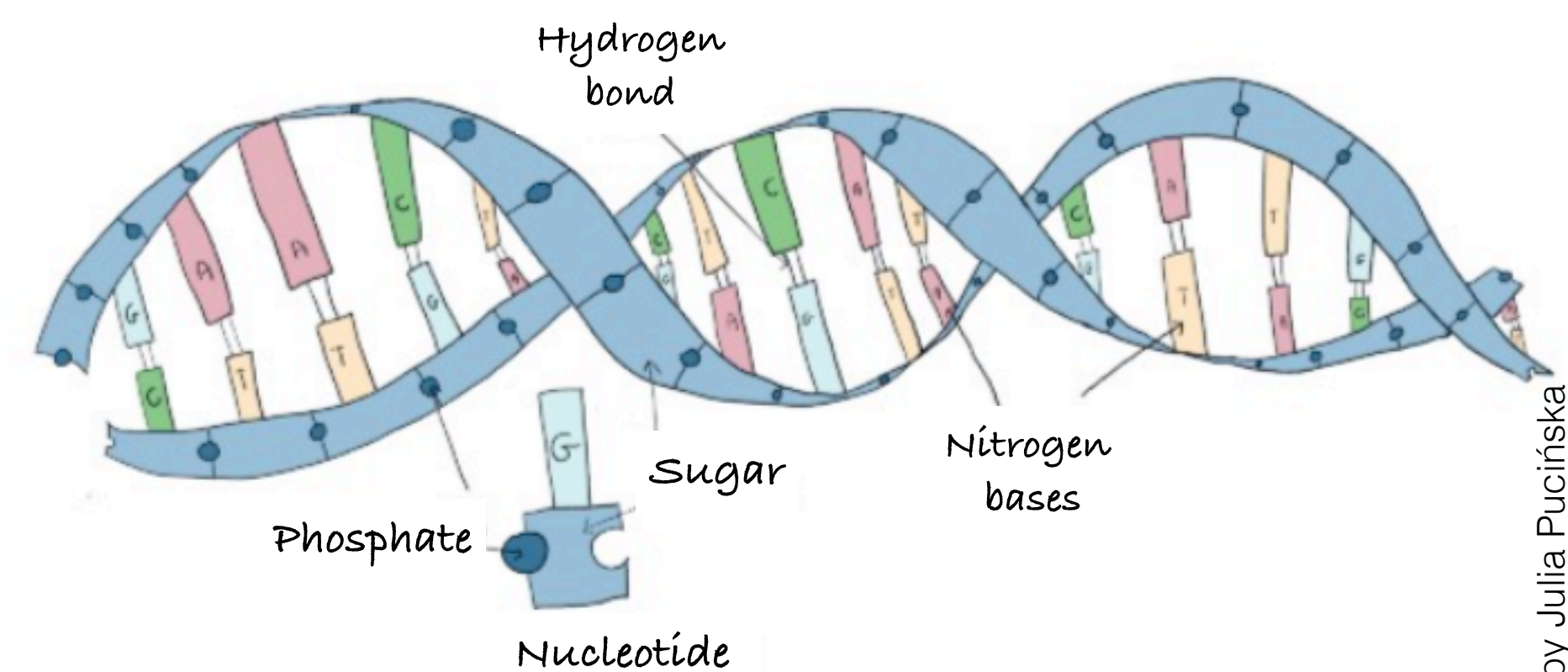


radiobiology

30-150 nm



a few nm



nanodosimetry

MC modeling

Laboratories



in silico



in vitro

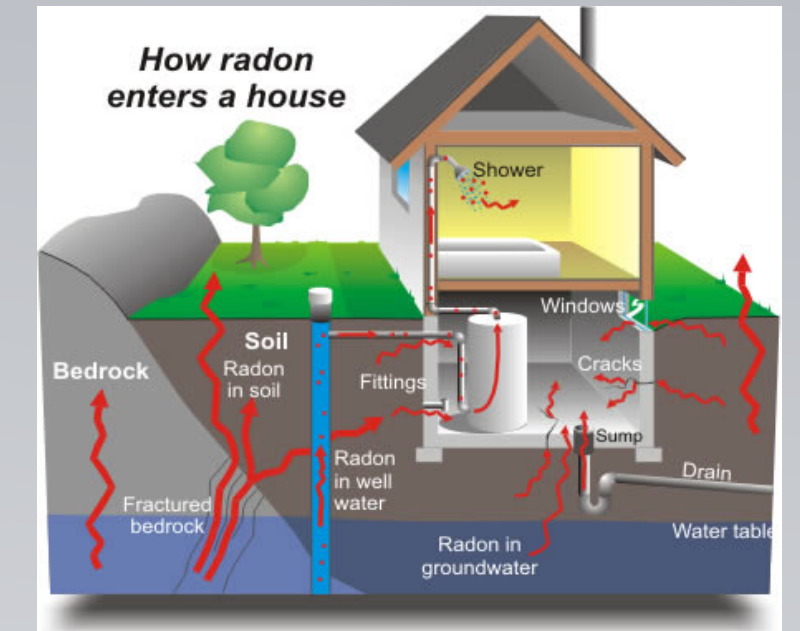


Outline

- Motivation: why mixed beams?
- DSB, SSB, 53BP1, GFP
- Focus analysis:
 - A. Damage induction
 - B. Repair kinetics
- Ongoing projects

Who is exposed to mixed beams of radiation?

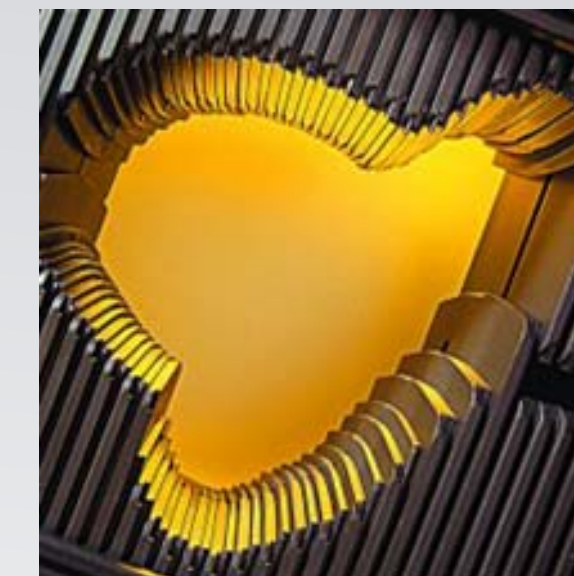
- People living in areas of high natural background radiation ($\alpha + \gamma$)



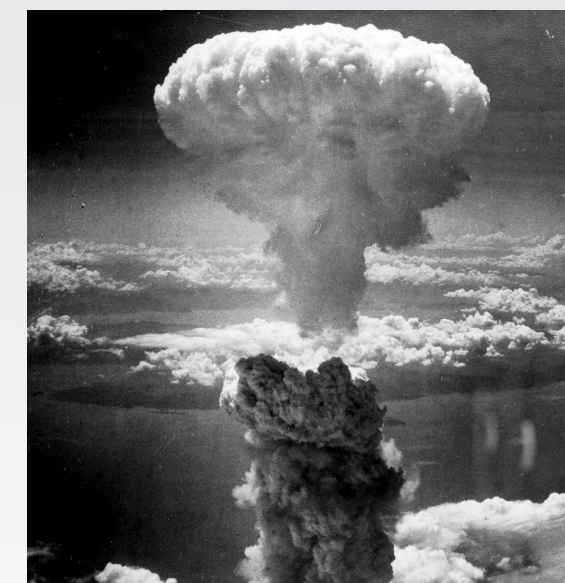
- Aeroplane passengers and astronauts ($n + p + \gamma$)



- Cancer patients treated with IMRT and proton therapy ($n + \gamma$)



- People involved in radiation accidents

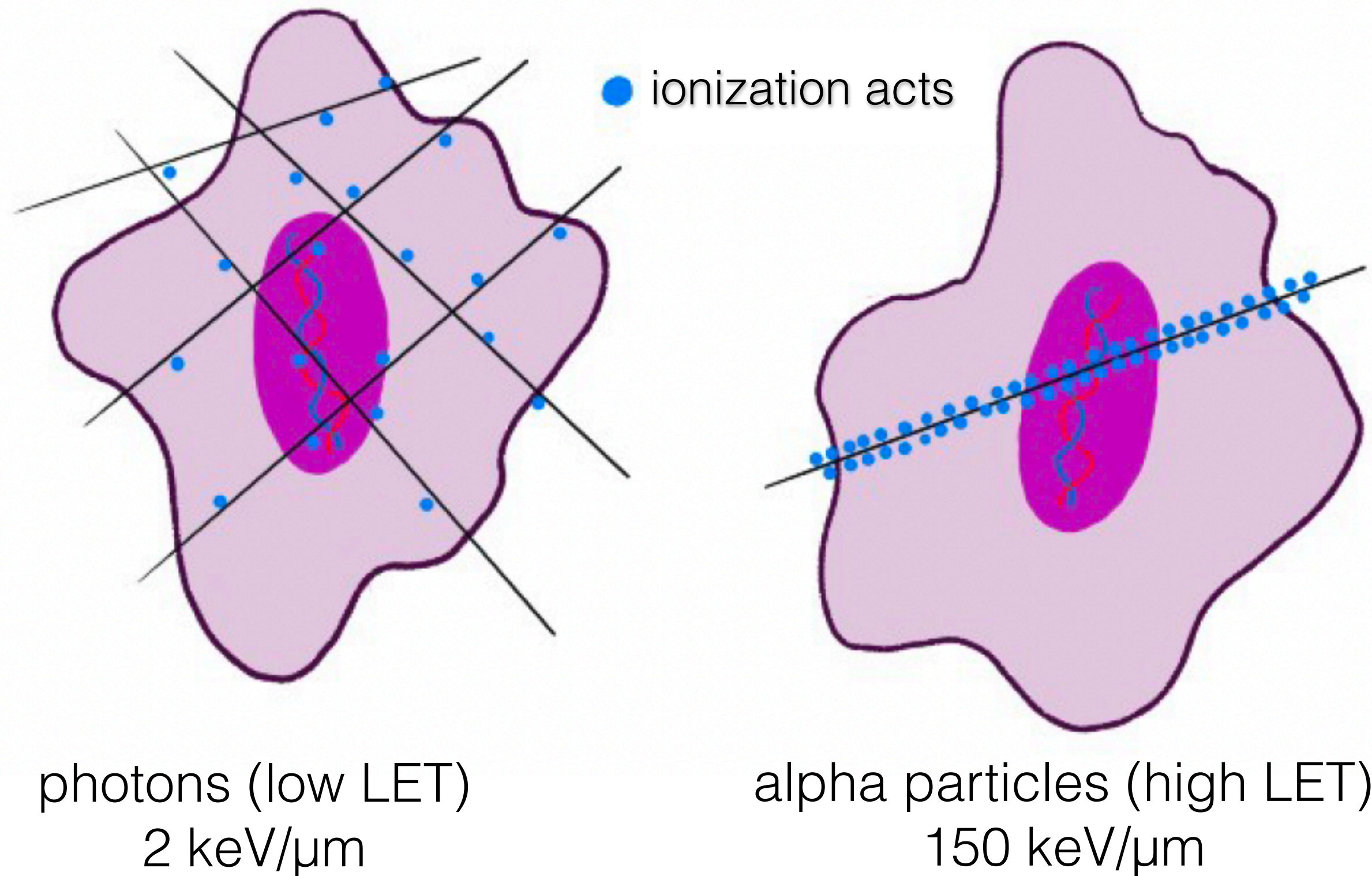


Biological effectiveness

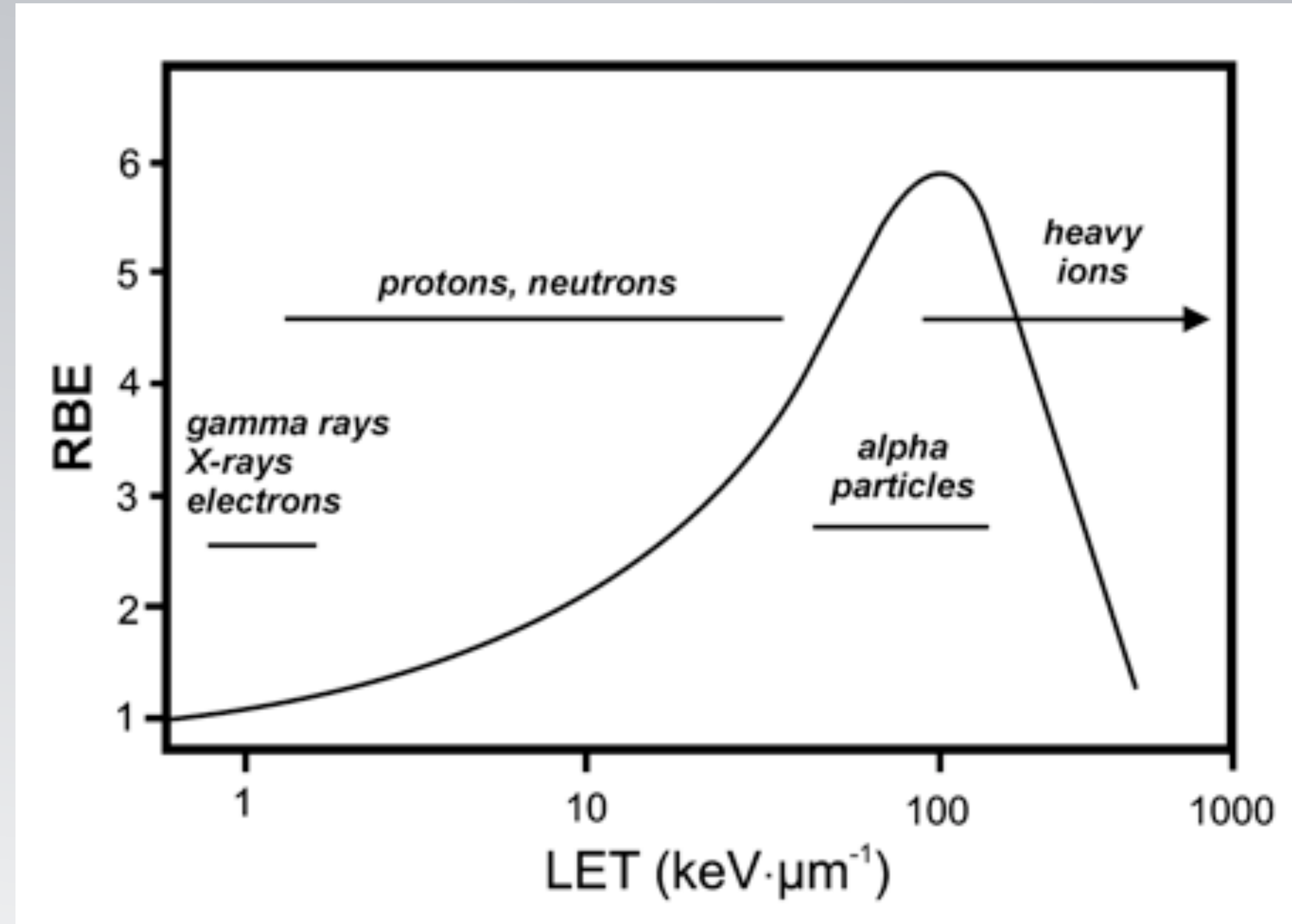
Absorbed dose: $D = dE/dm$

RBE - Relative Biological Effectiveness

courtesy of J. Pucinska



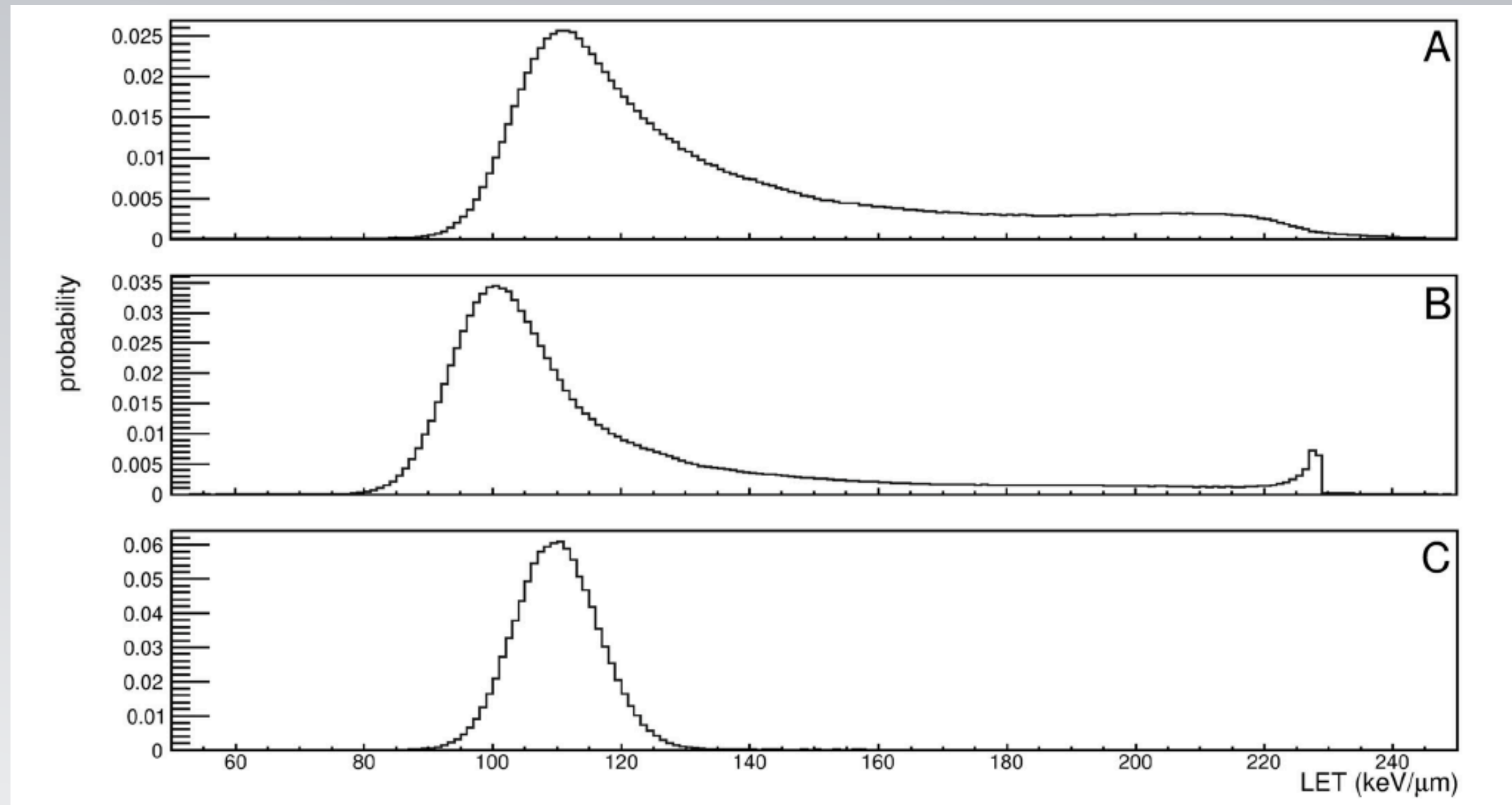
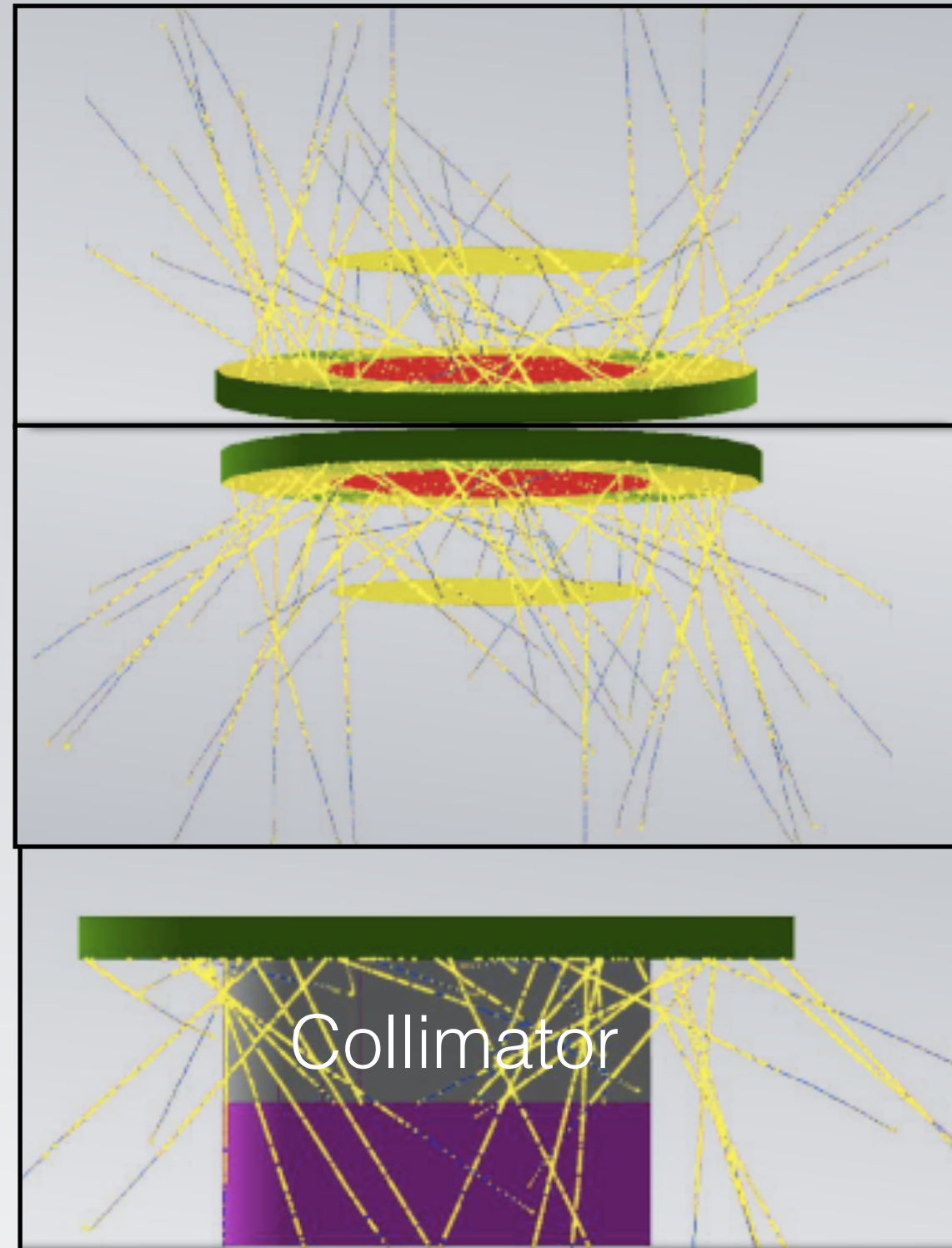
LET (Linear Energy Transfer) = dE/dx



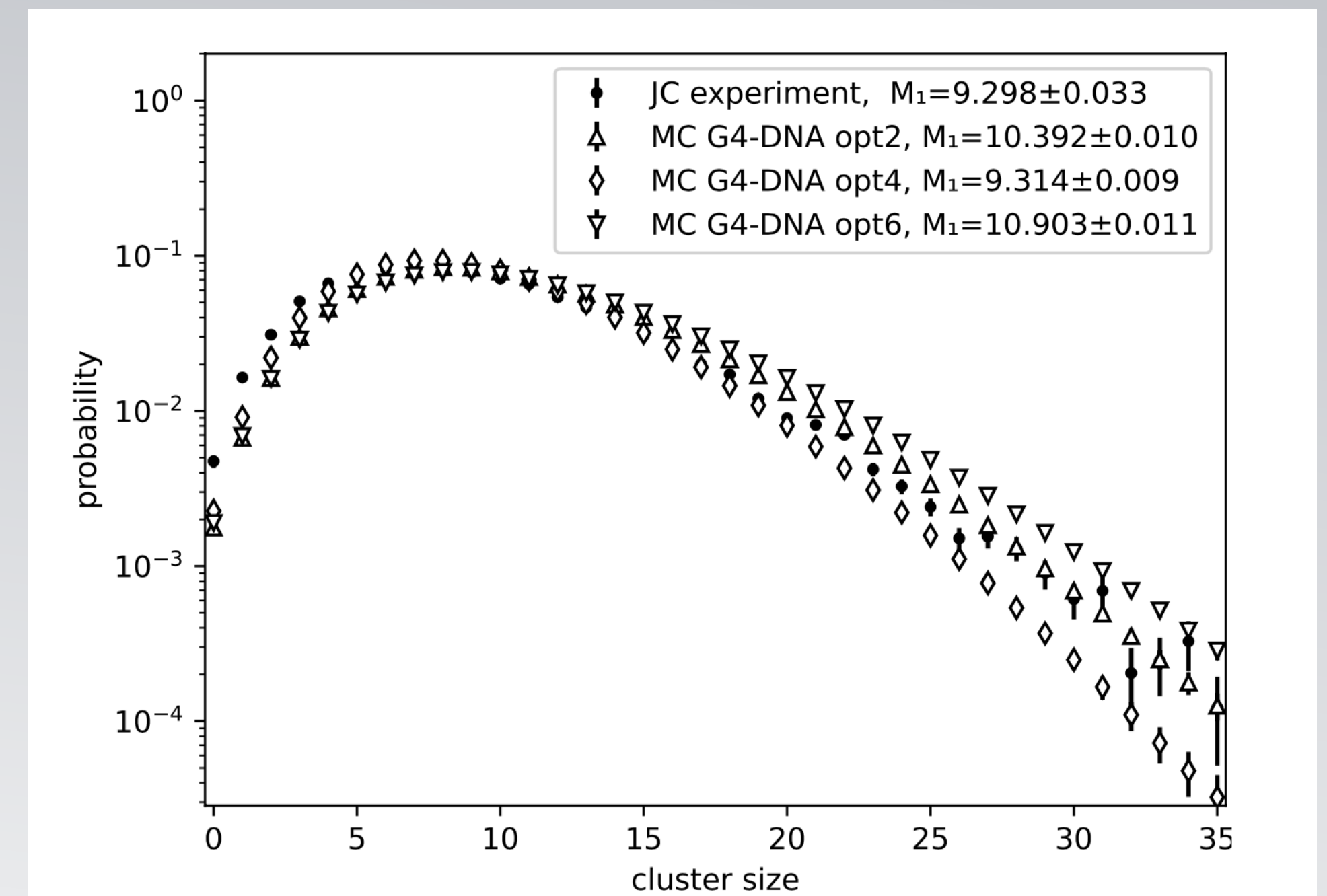
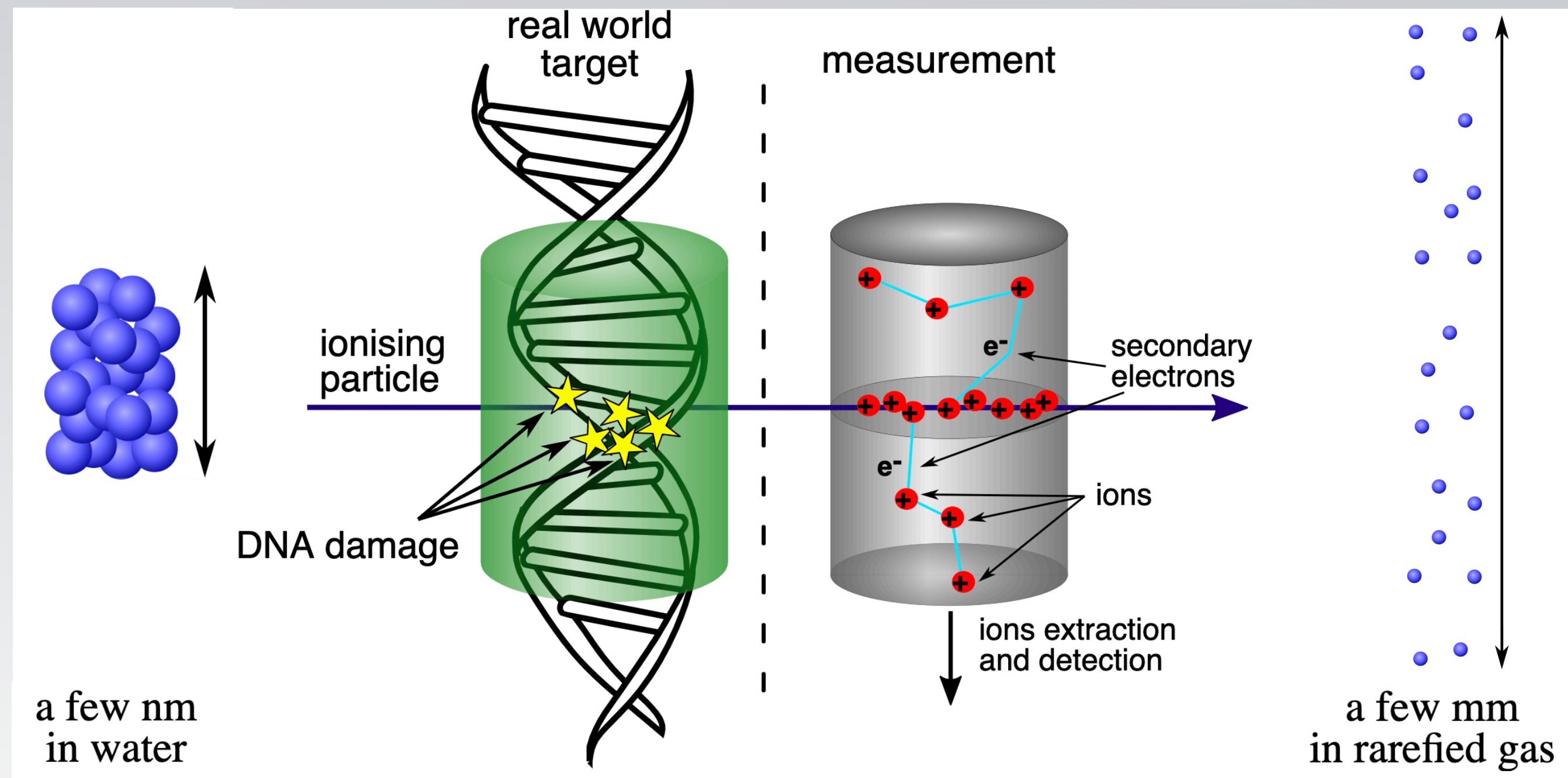
$RBE_{50\%SF} = D_{\text{ref}}/D_{\text{test}}$

LET distributions

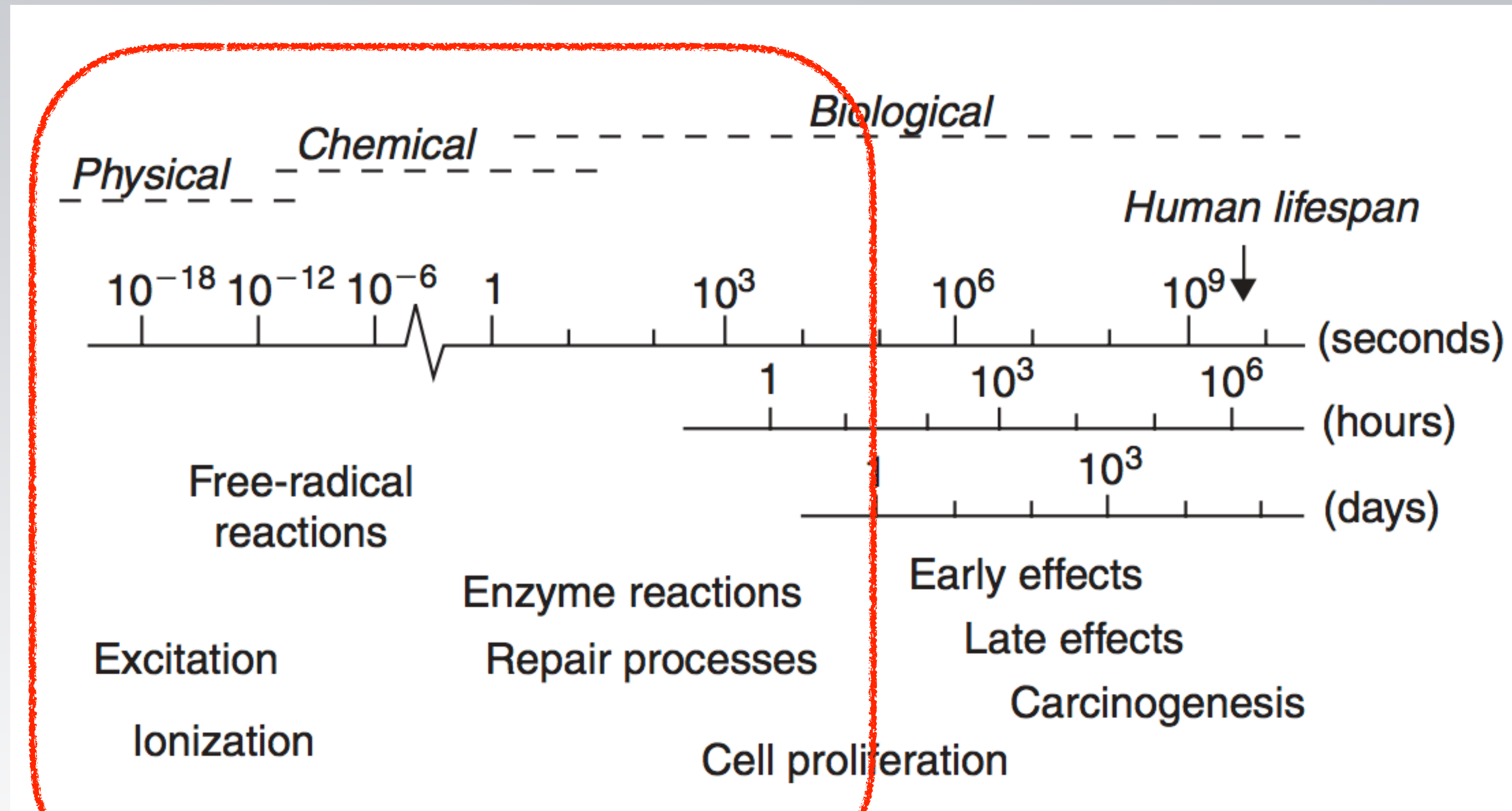
^{241}Am



Nanodosimetry: measurements of ionisations



Radiation exposure on biological systems



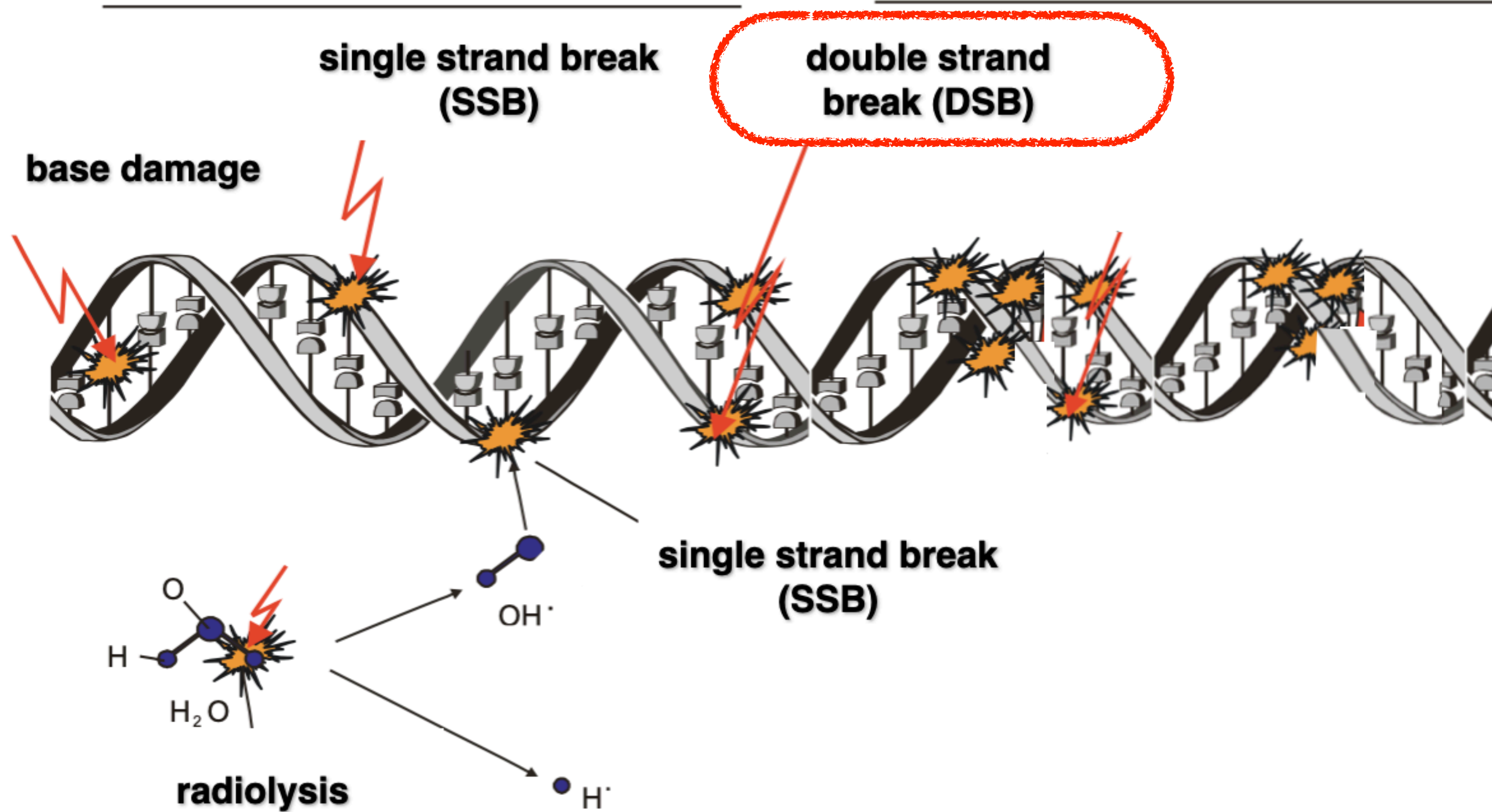
DNA damage

low LET

high LET

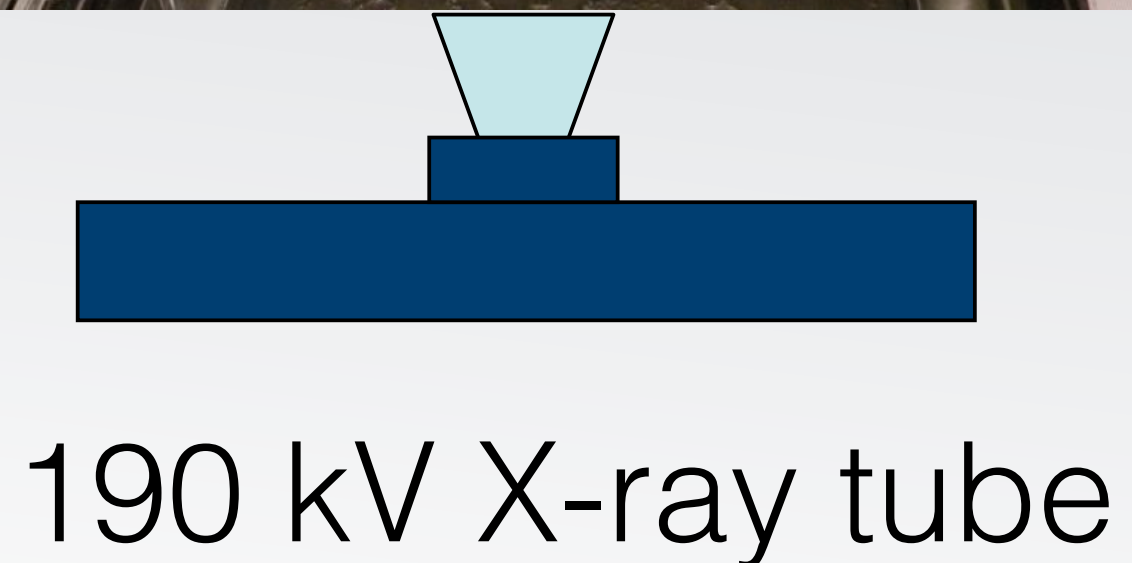
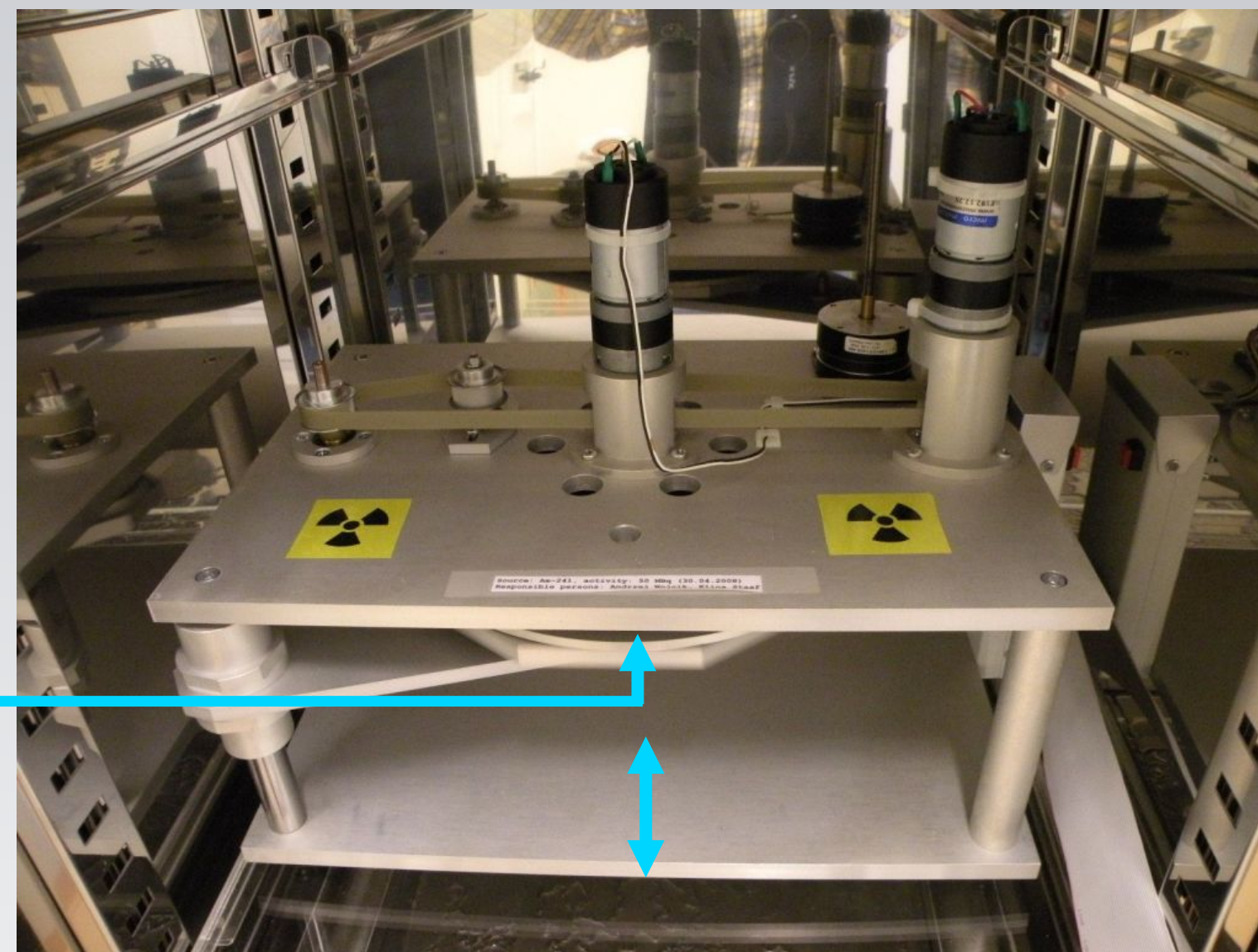
X-rays, gamma, beta radiation

alpha particles, protons, neutrons, etc.



Mixed beam exposure facility at Stockholm University

^{241}Am (0.21 Gy/min)

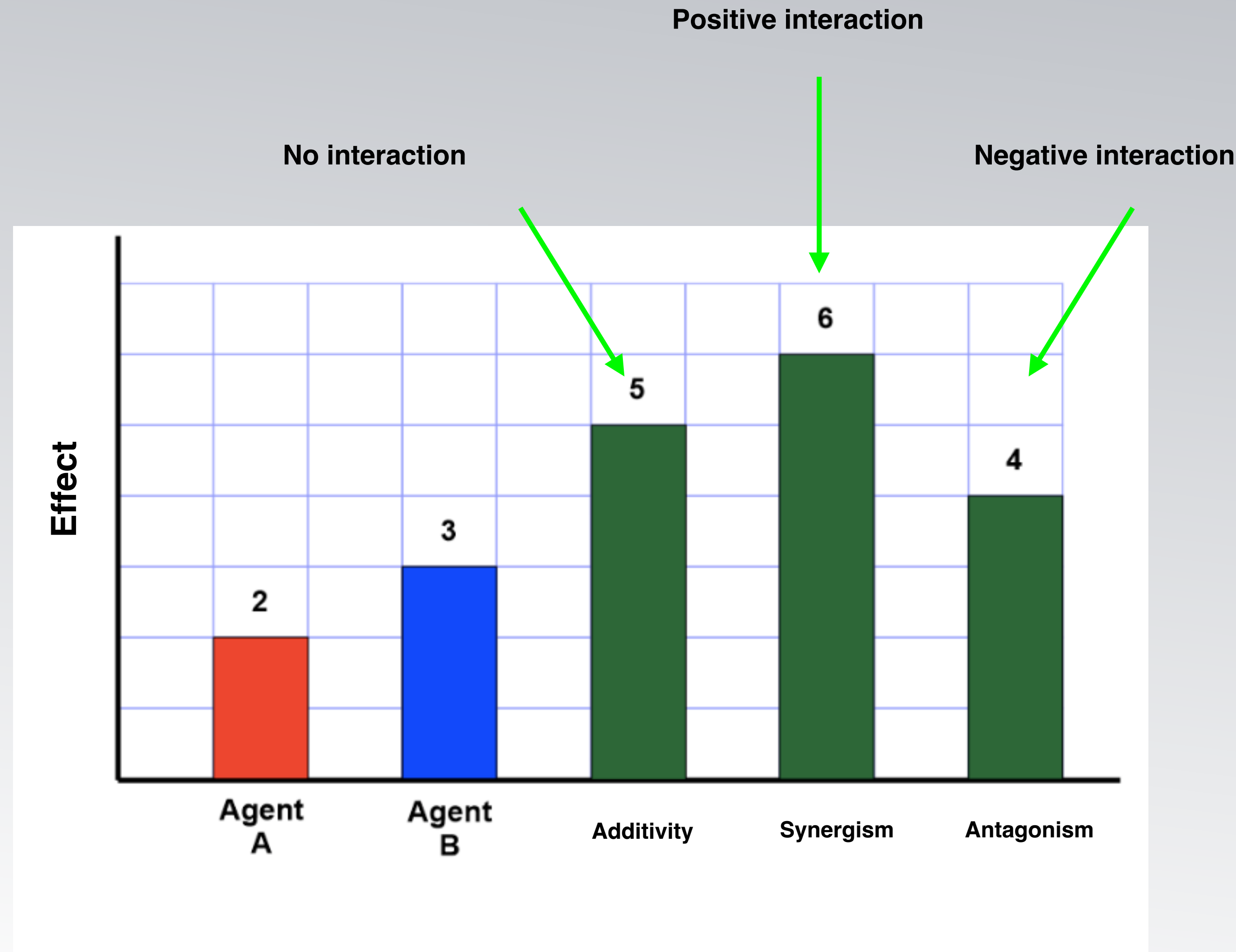


190 kV X-ray tube

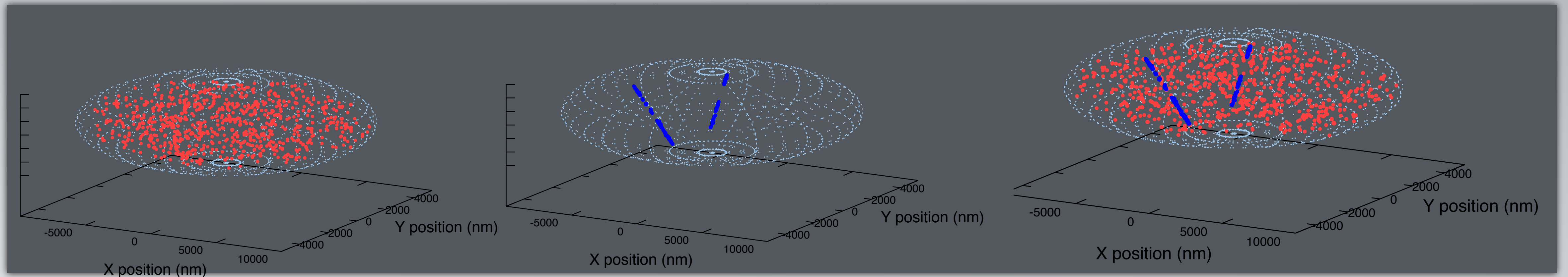
A schematic diagram showing a blue rectangular block representing the X-ray tube, with a blue trapezoidal shape representing the beam area above it. A blue double-headed arrow indicates the distance between the X-ray tube and the petri dish.

0.05 Gy/min
↕
0.07 Gy/min

Synergism or additivity?



Mixed beam irradiation



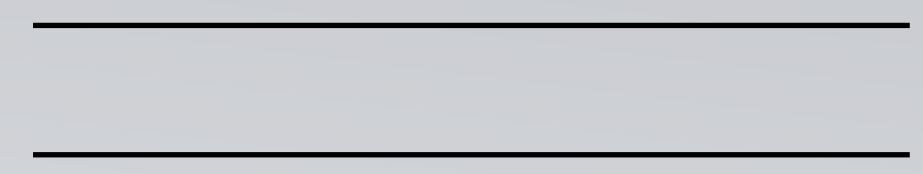
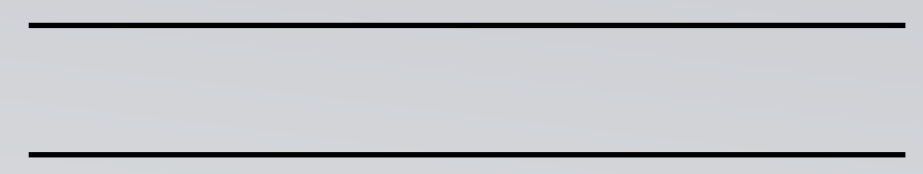
photons (0.25 Gy) + alpha particles (0.25 Gy) = mixed beams (0.5 Gy)

Source	Dose (Gy)	DSB	SSB	SSB/DSB
α particles	0.235	25.8	150.5	5.8
X-rays	0.253	13.5	241.3	17.9
Mixed beams	0.488	38.9	391.7	10.06

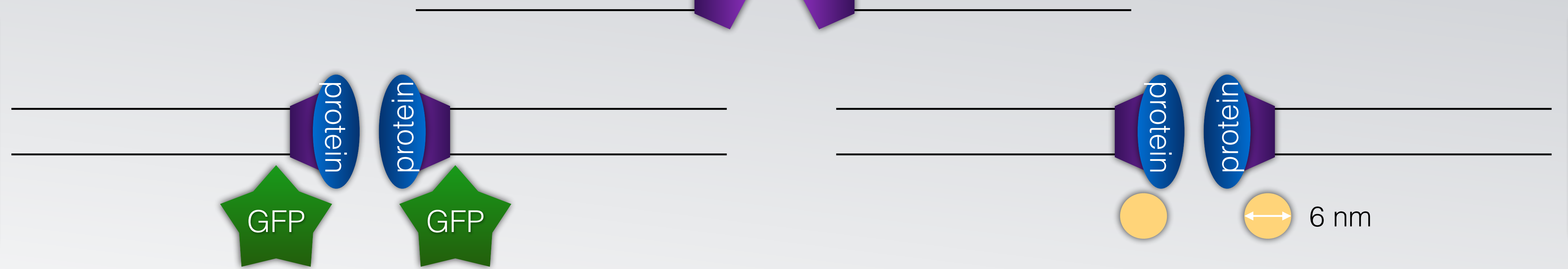
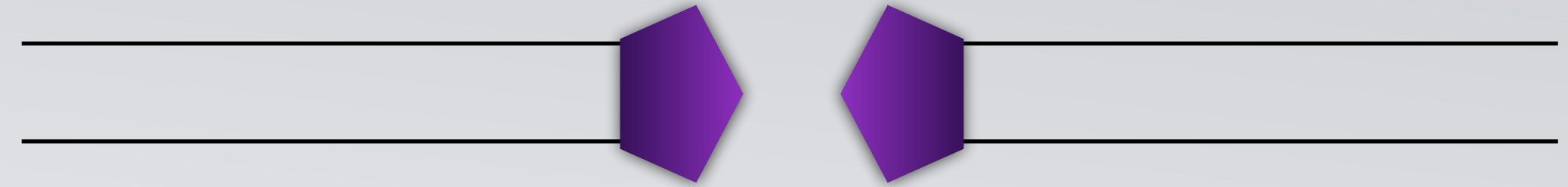
MC simulations with PARTRAC

How to visualize DSB?

double strand break



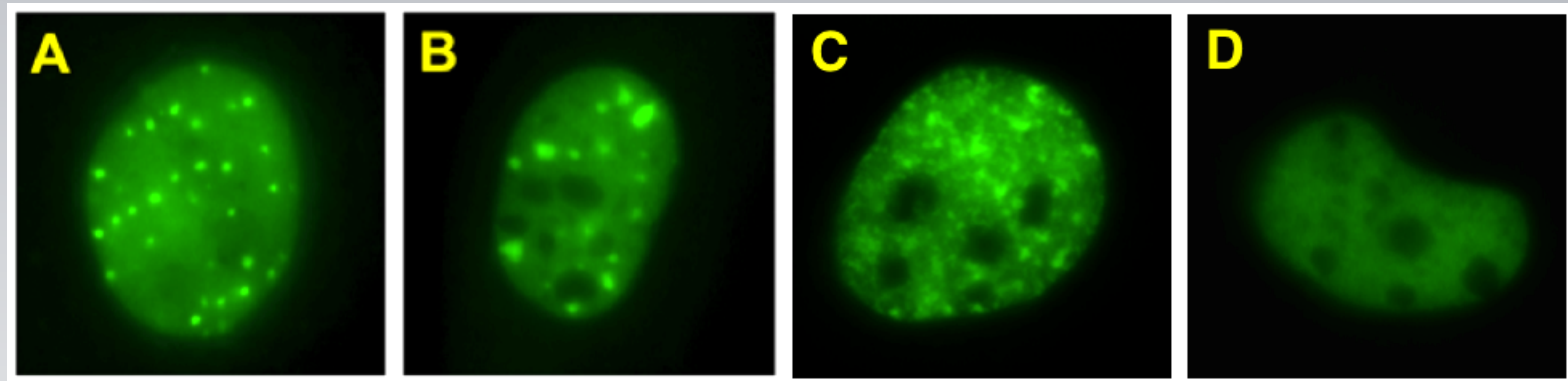
}DNA strand



focus analysis

TEM analysis

Radiation-induced foci 53BP1

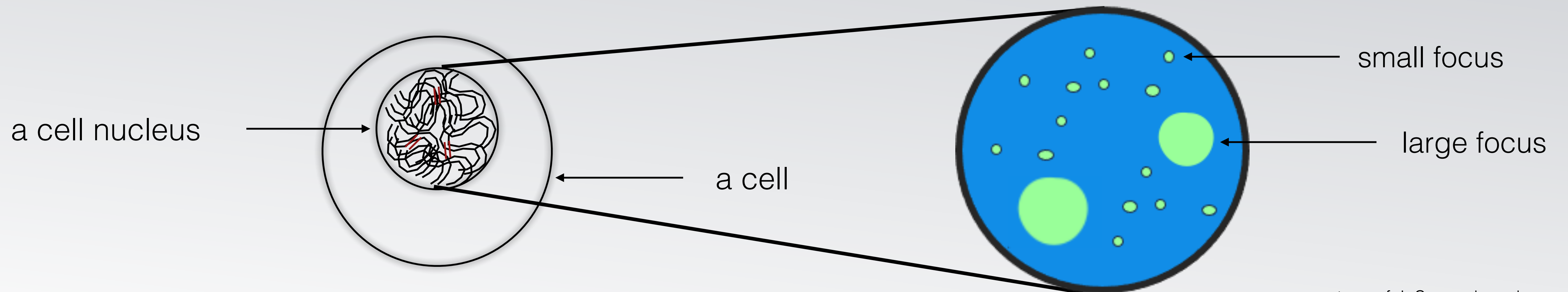


X-rays

alpha particles

mixed beams

control cells (not irradiated)

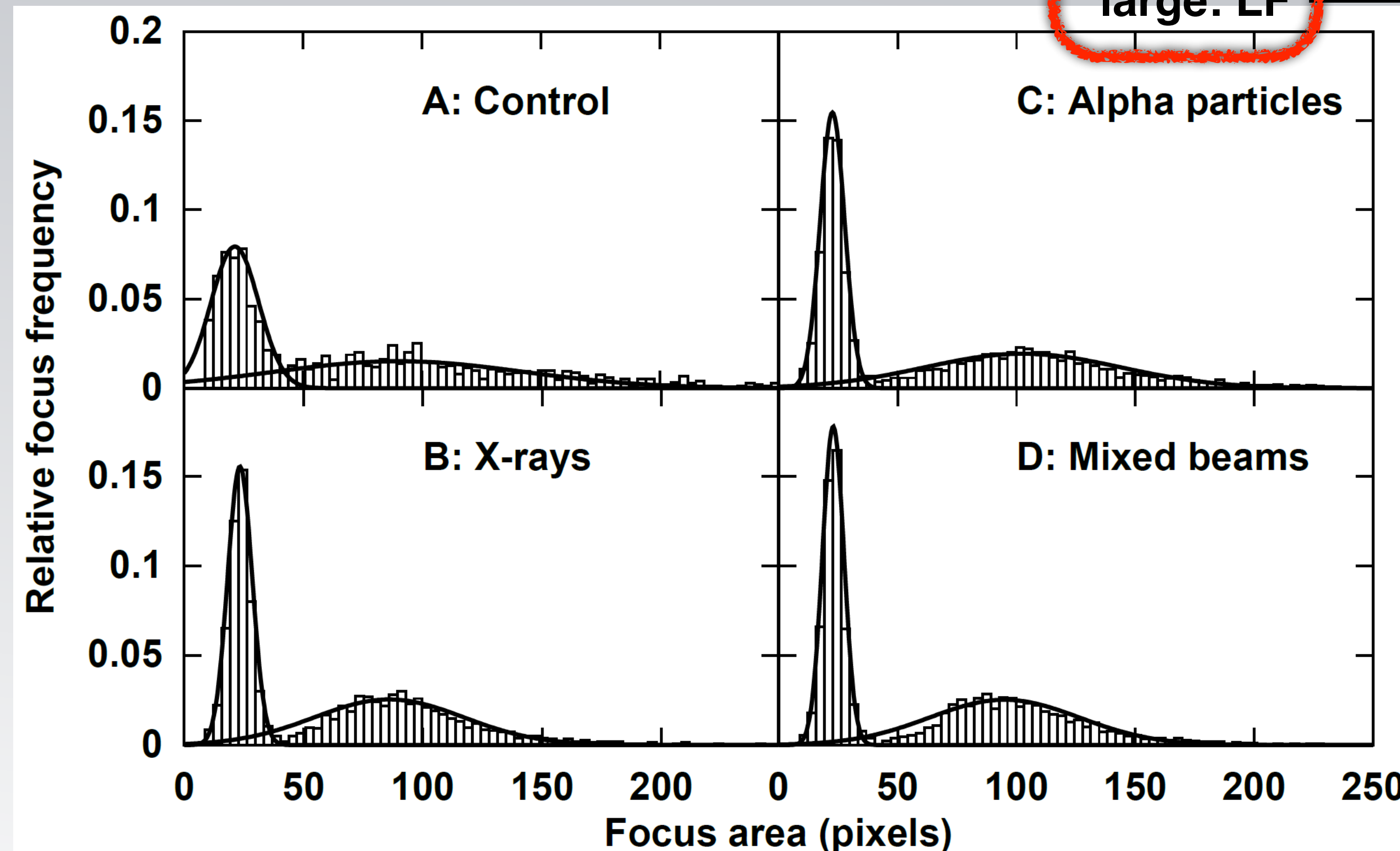
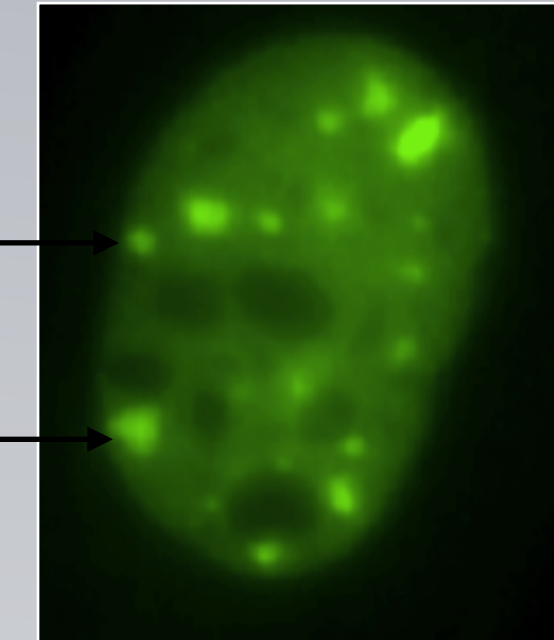


courtesy of J. Szczechowska

Focus size

AF

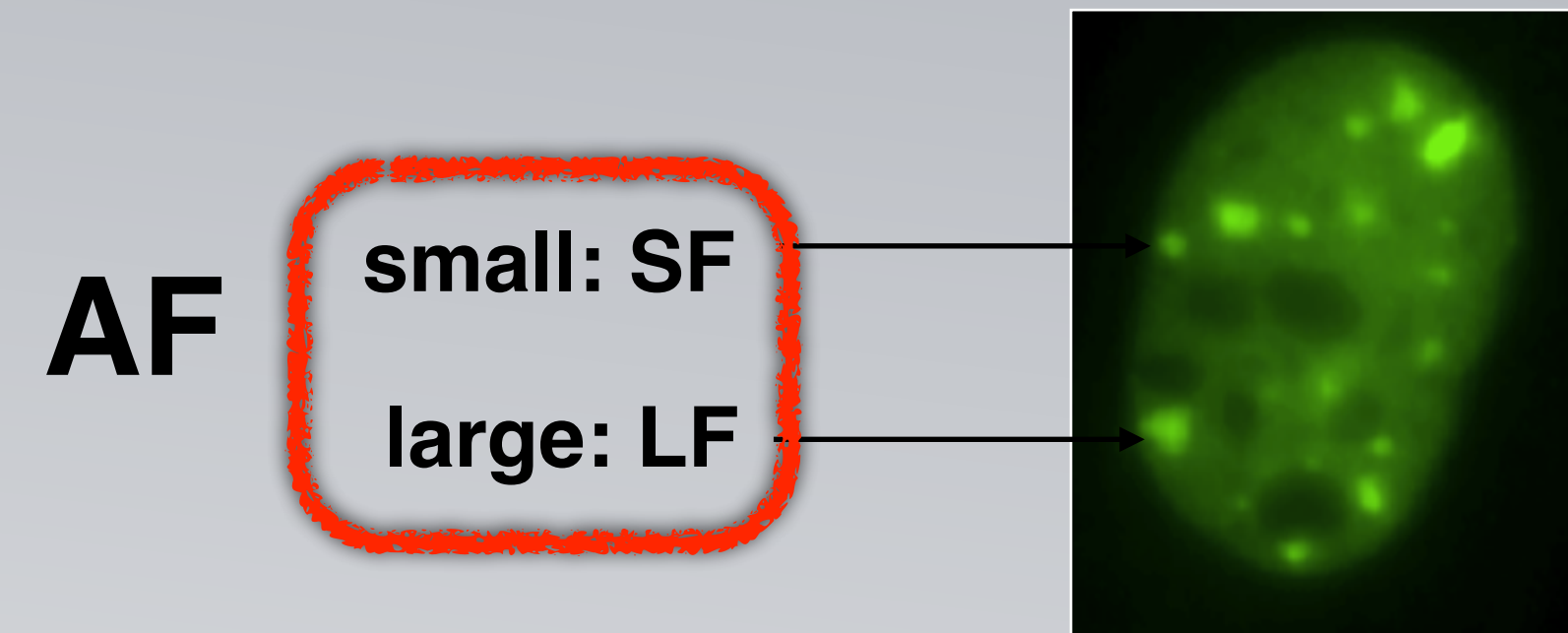
small: SF
large: LF



SF - likely containing simple DSB

LF - likely containing staggered complex DSB

Focus analysis



STEP 1: cell response on radiation - focus frequency as a function of dose measured 0.5 h after exposure

STEP 2: repair kinetics - focus frequency as a function of time for the dose equal 1 Gy

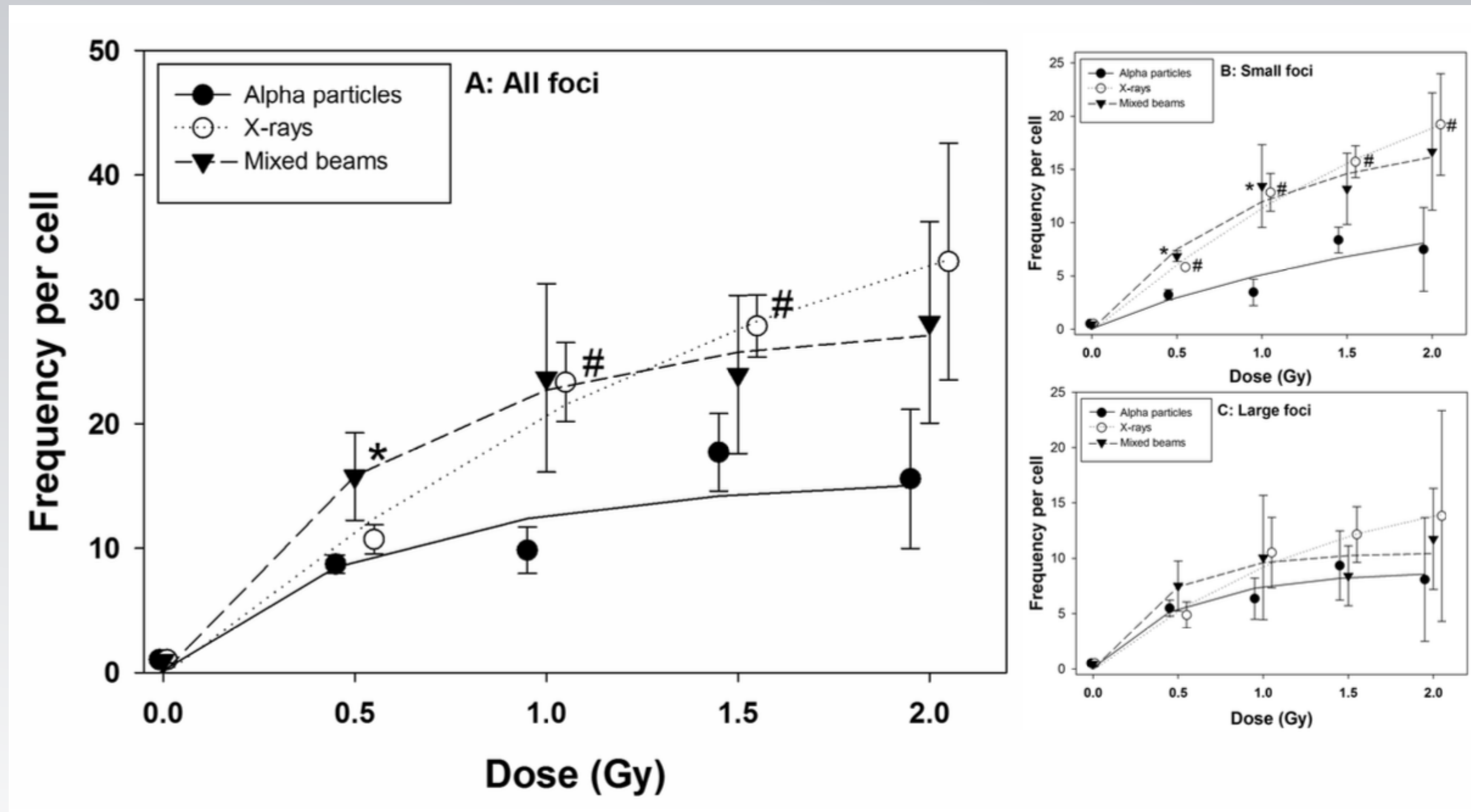
Dose response

STEP 1: cell response on radiation - focus frequency as a function of dose measured 0.5 h after exposure



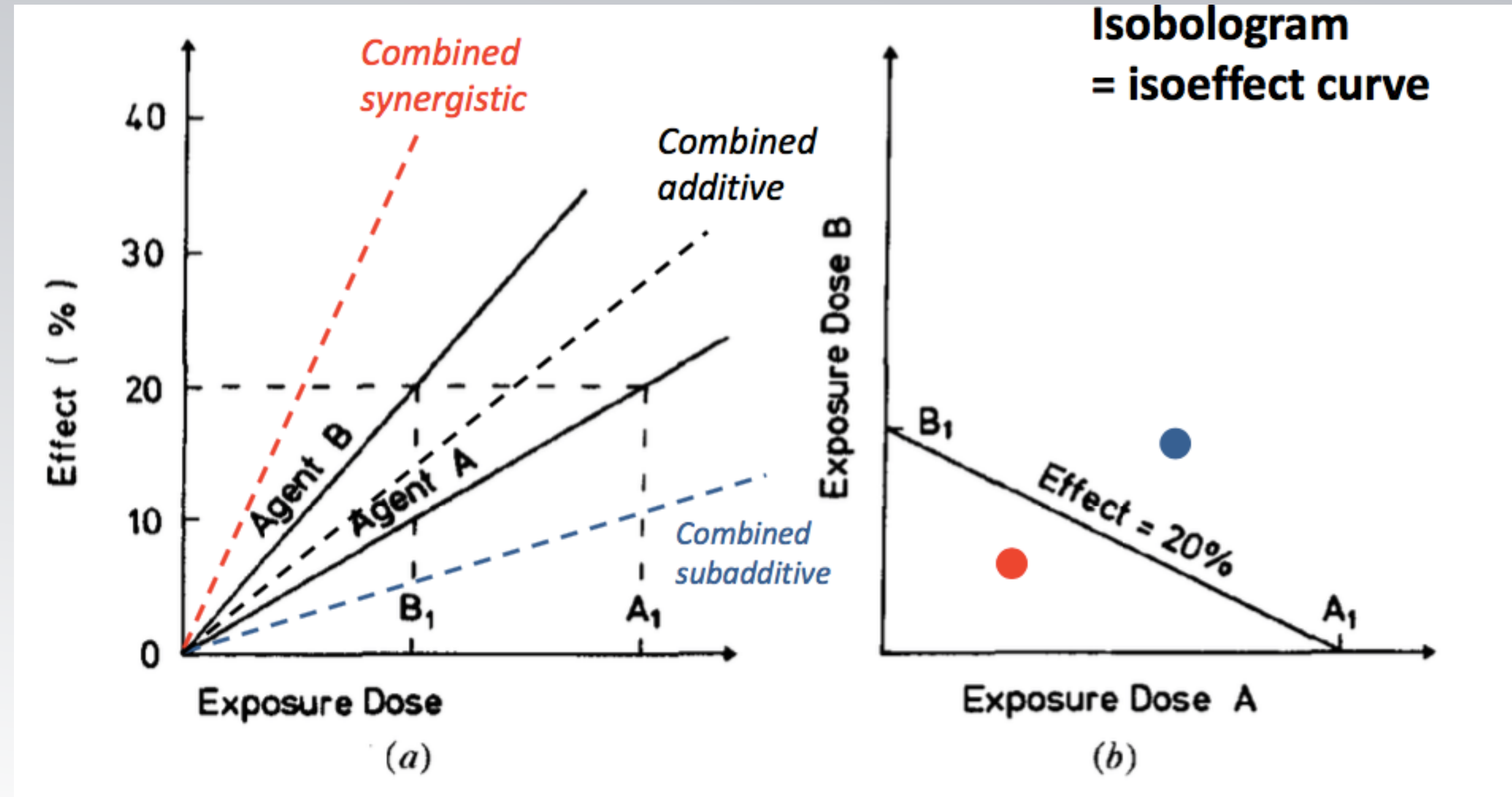
t = 0.5 h because the highest focus frequencies

Dose response in U2OS cells

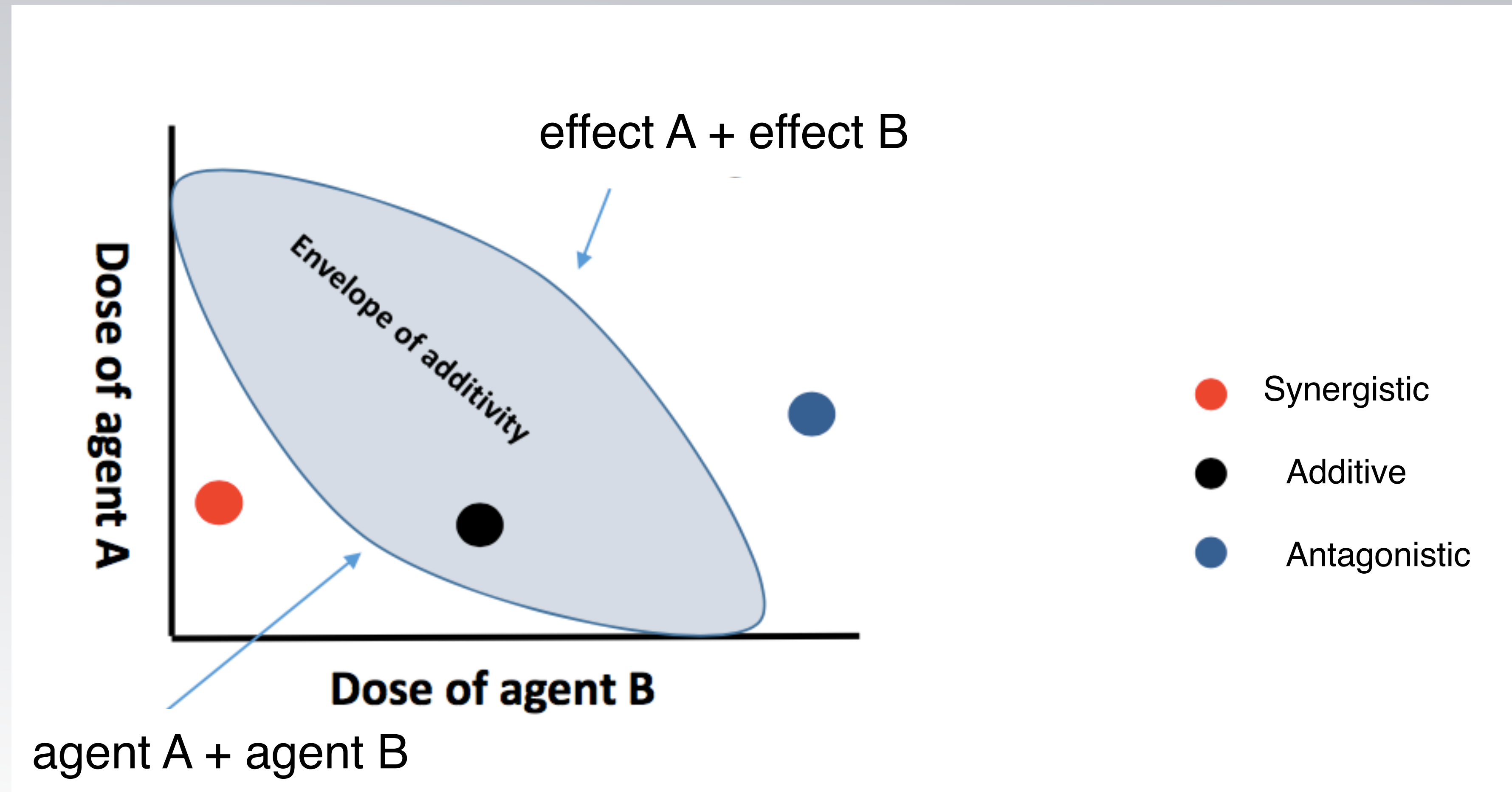


saturation: focus confluency
alpha particles: lowest frequencies of foci (AF, SF)

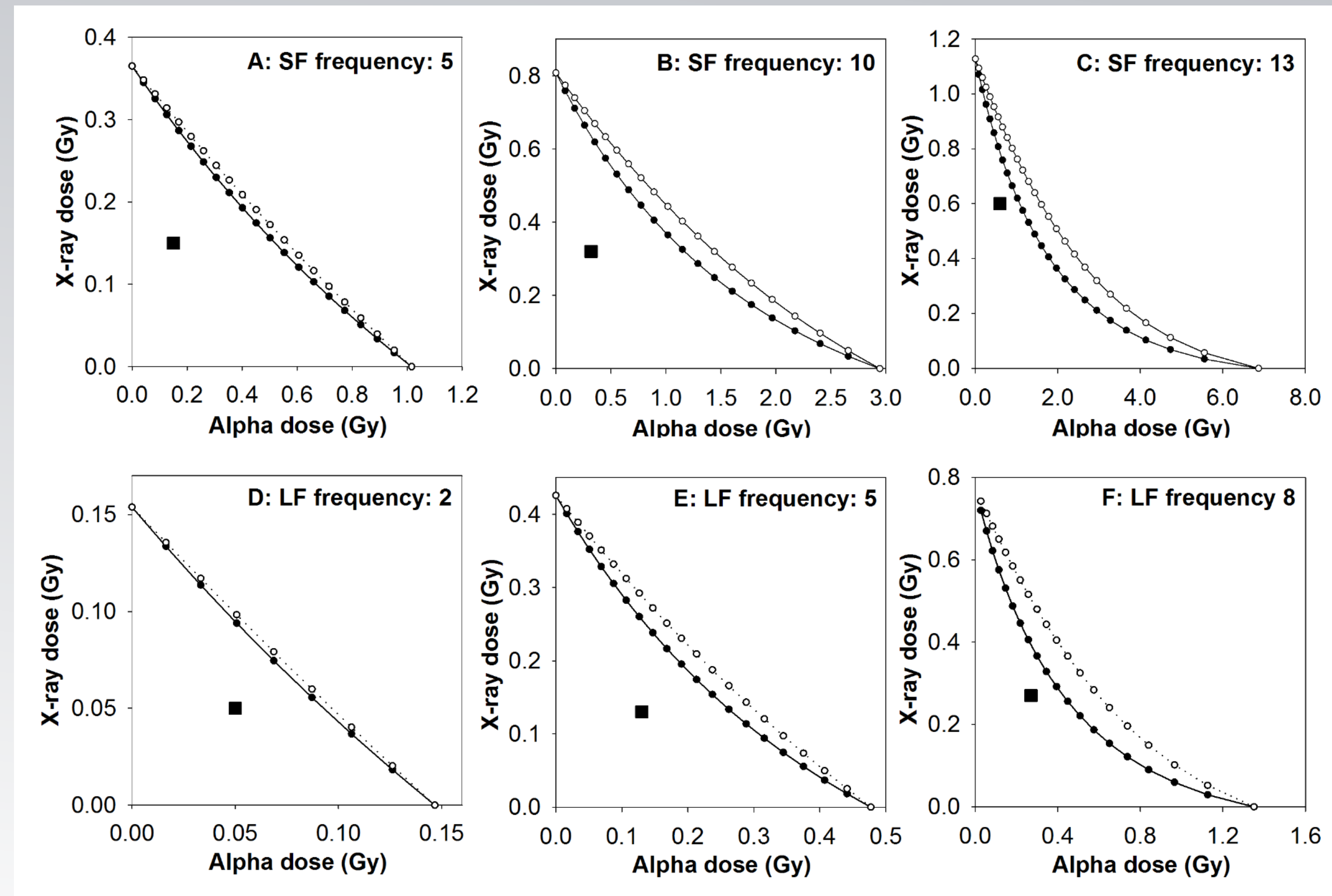
If the relationship is linear



If the dose response is **not** linear

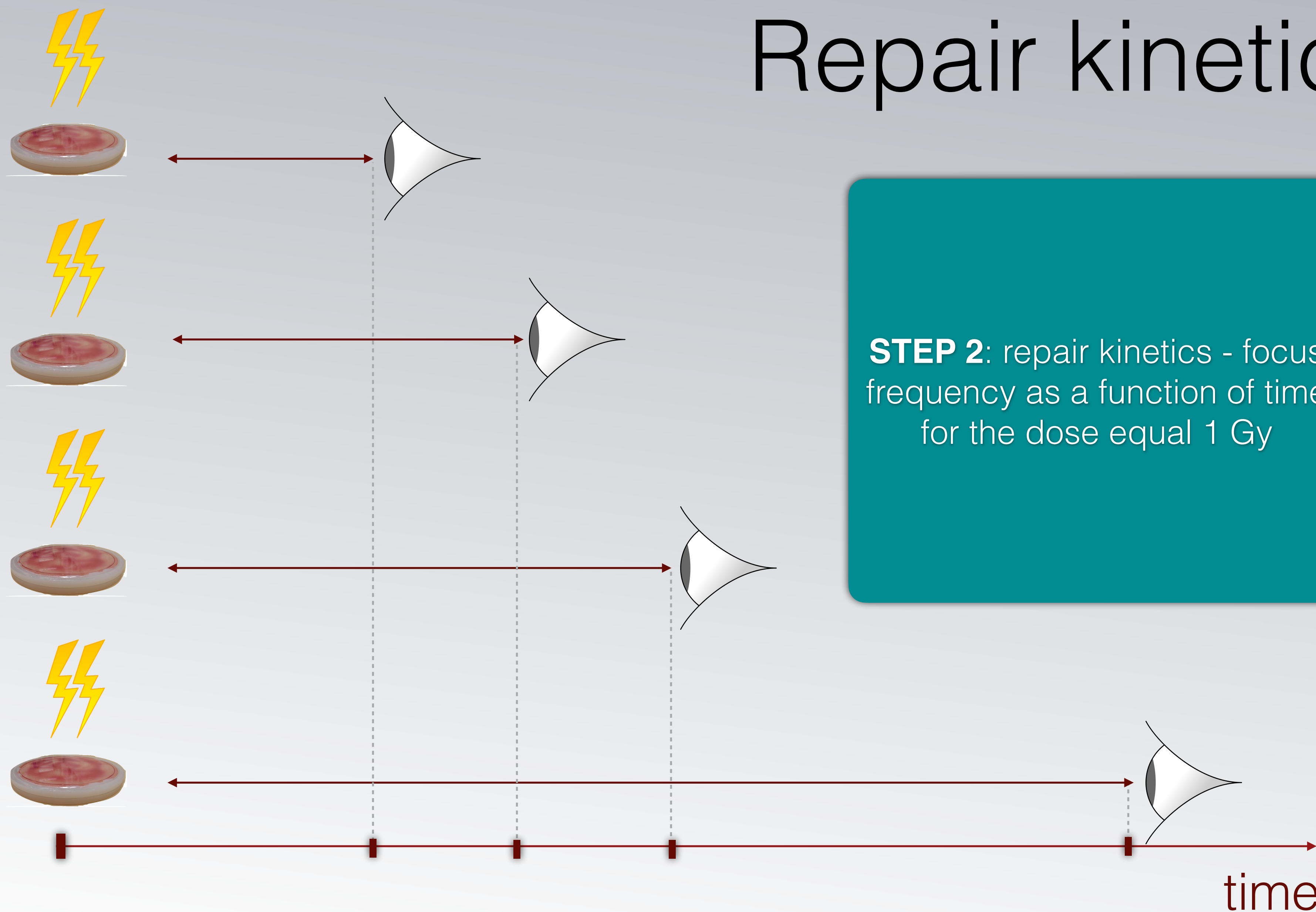


Envelopes of additivity for SF and LF

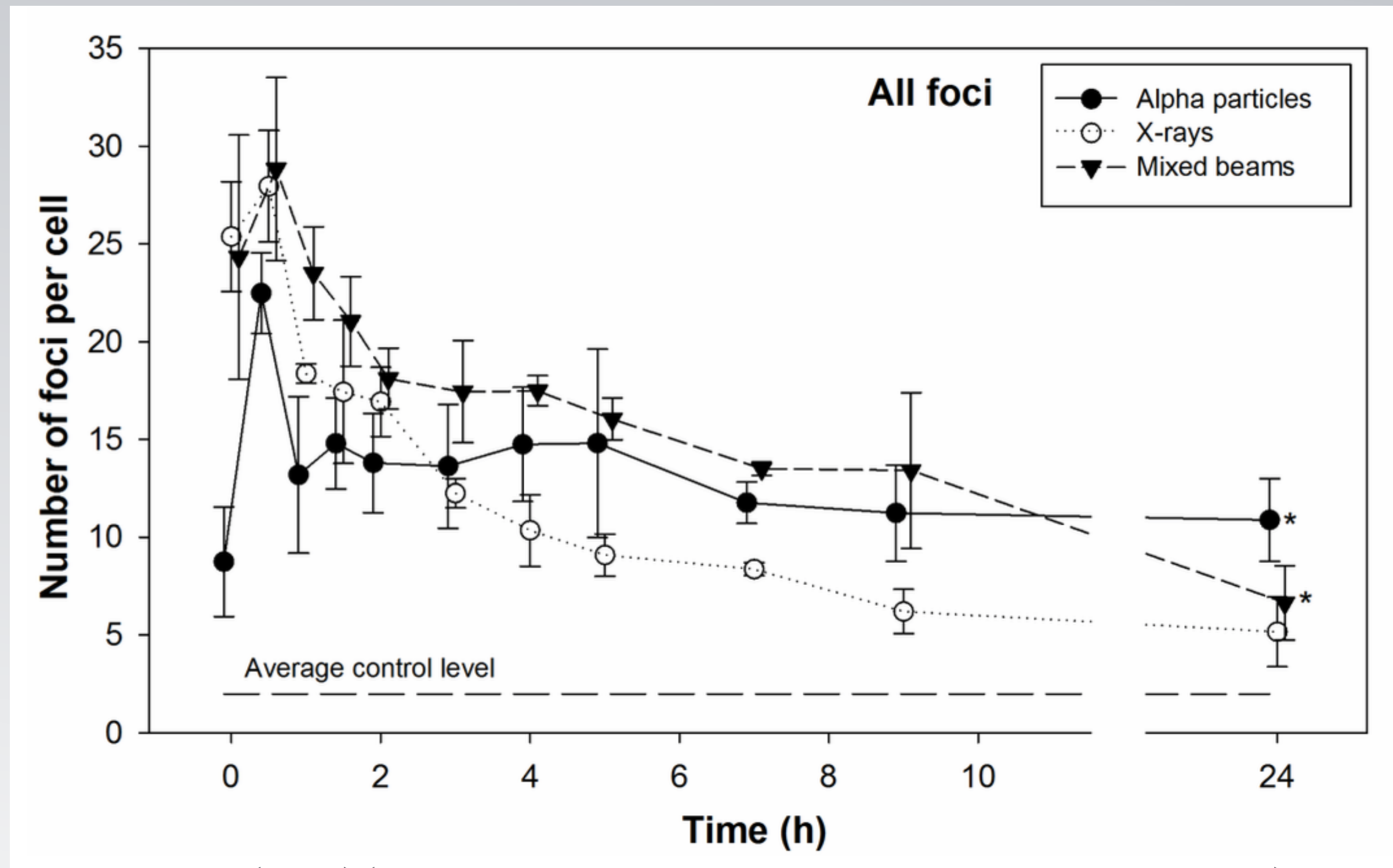


level of interaction is inversely related to the level of damage

Repair kinetics



Focus decay



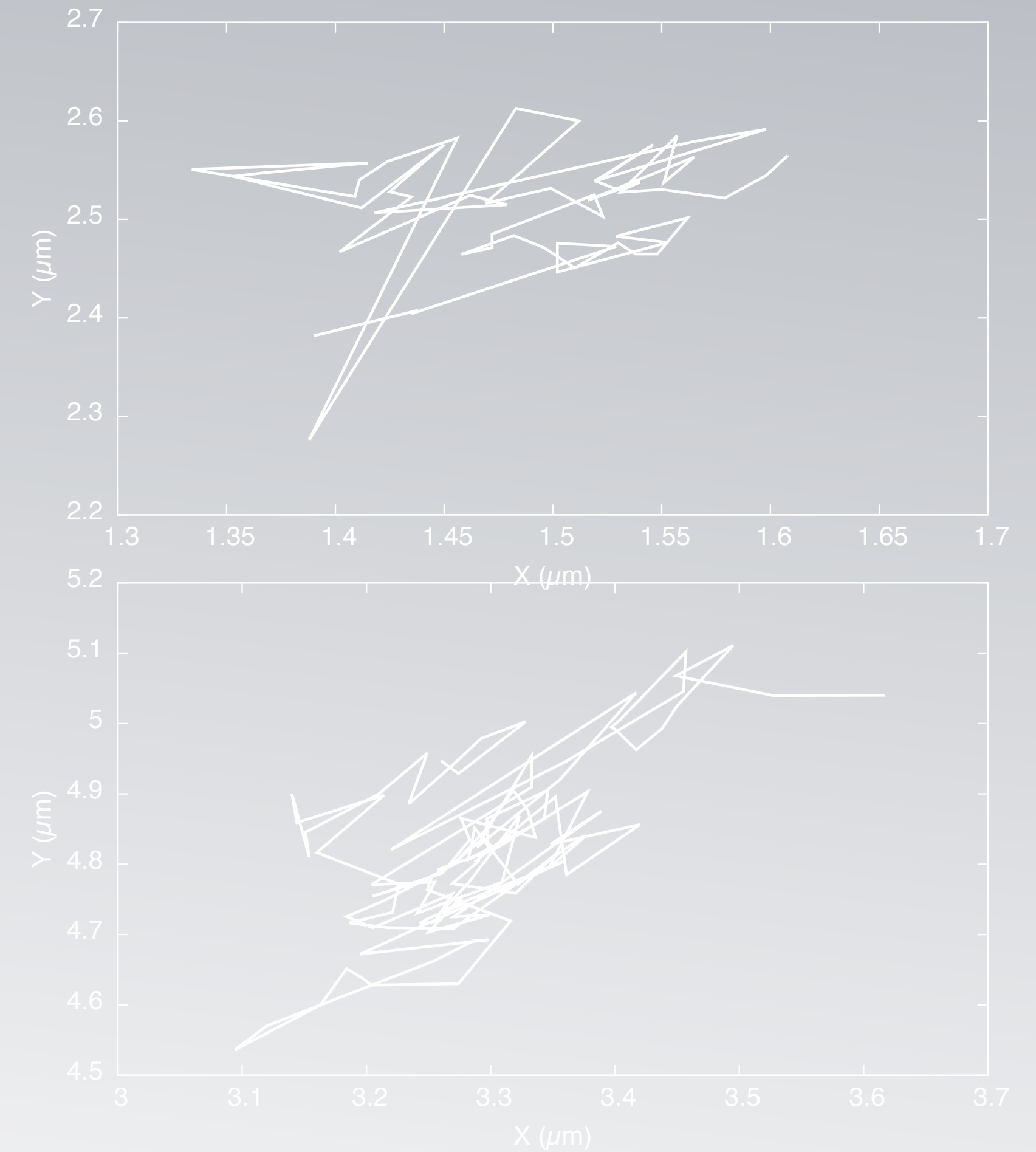
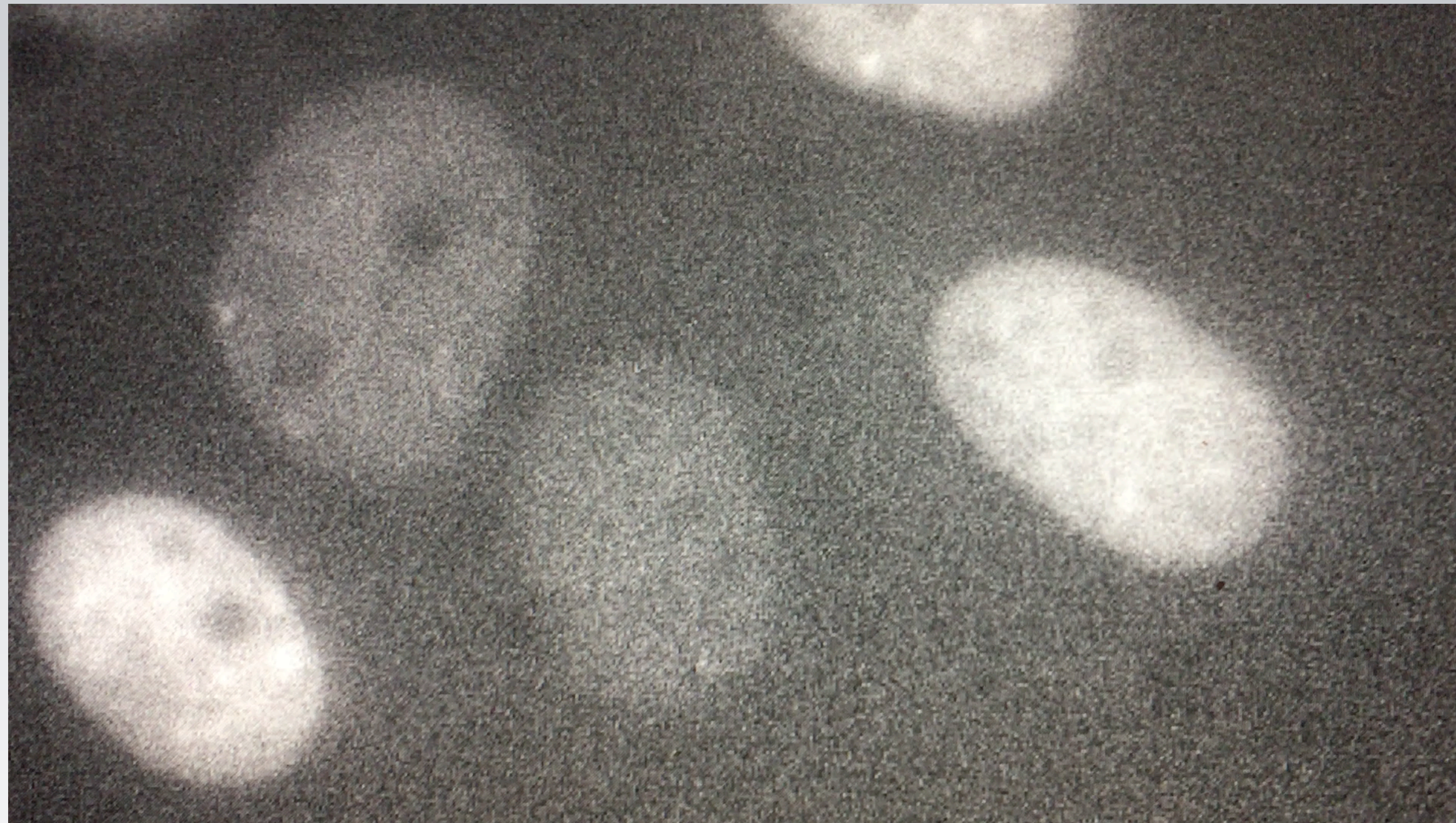
- focus appearance immediately after irradiation
- lower no. of foci for alpha particles at 30 min
- sharp decrease between 30 min and 1 h for alphas and X-rays (41%, 34%)
- slower decrease for mixed beams (18%)

Fast repair

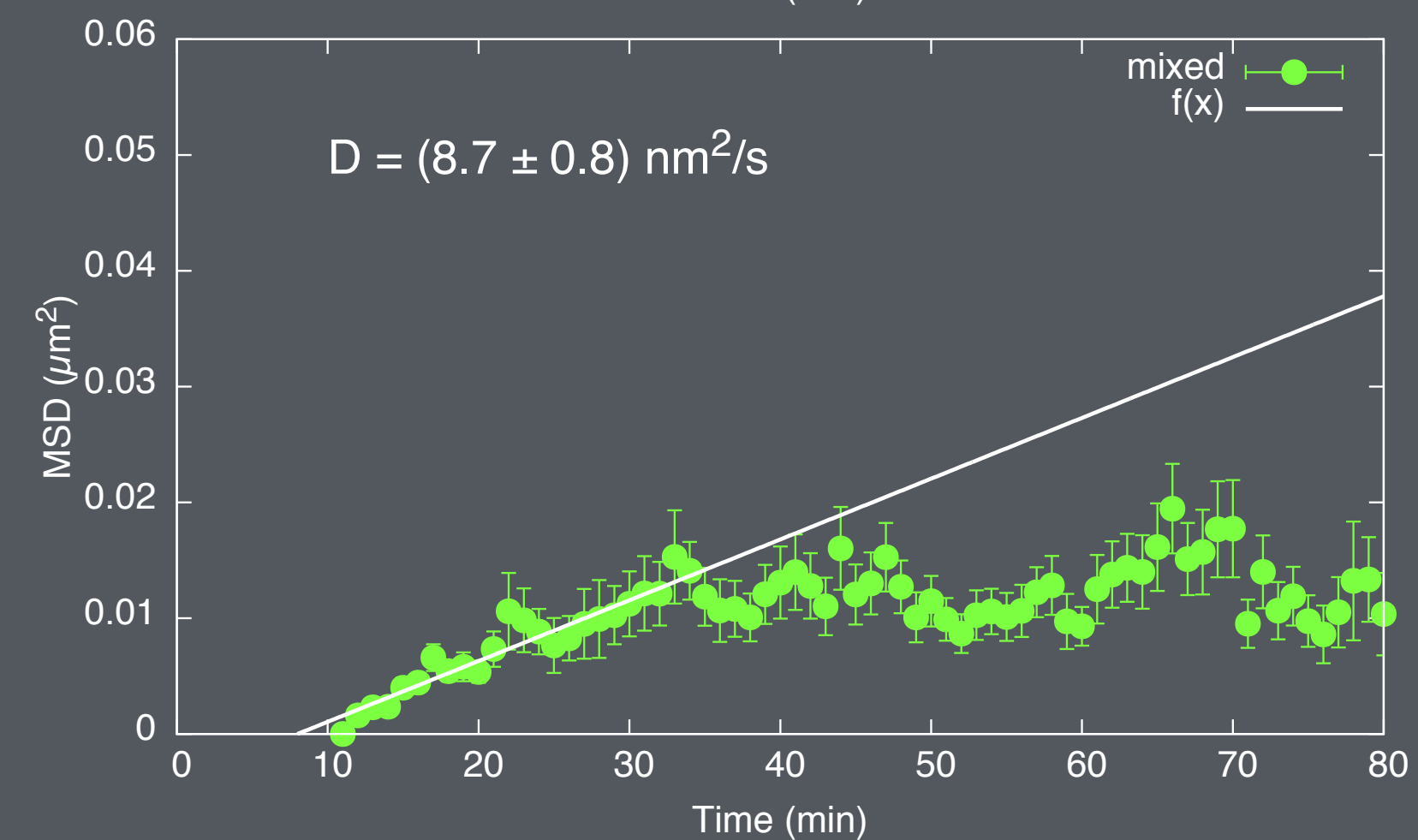
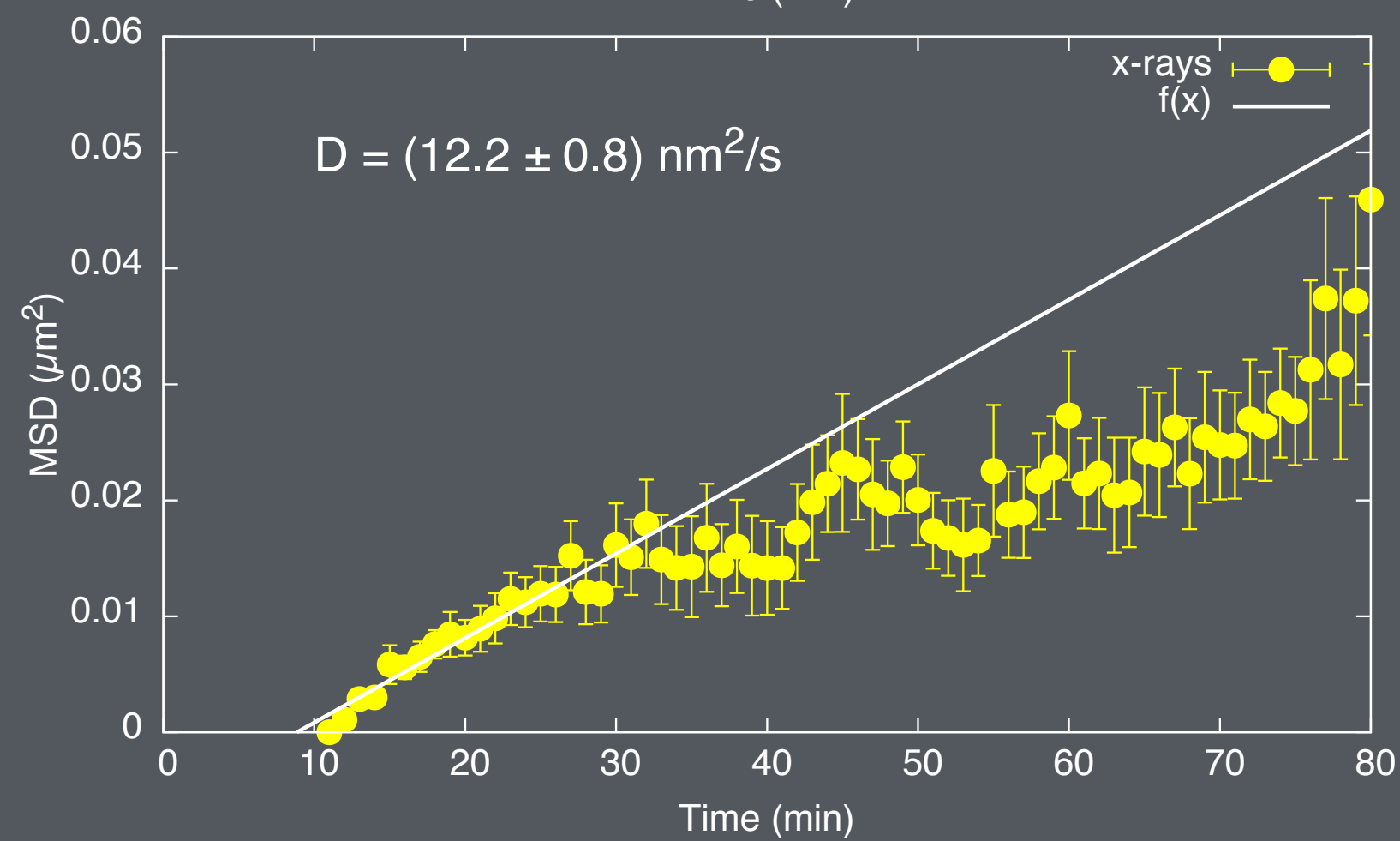
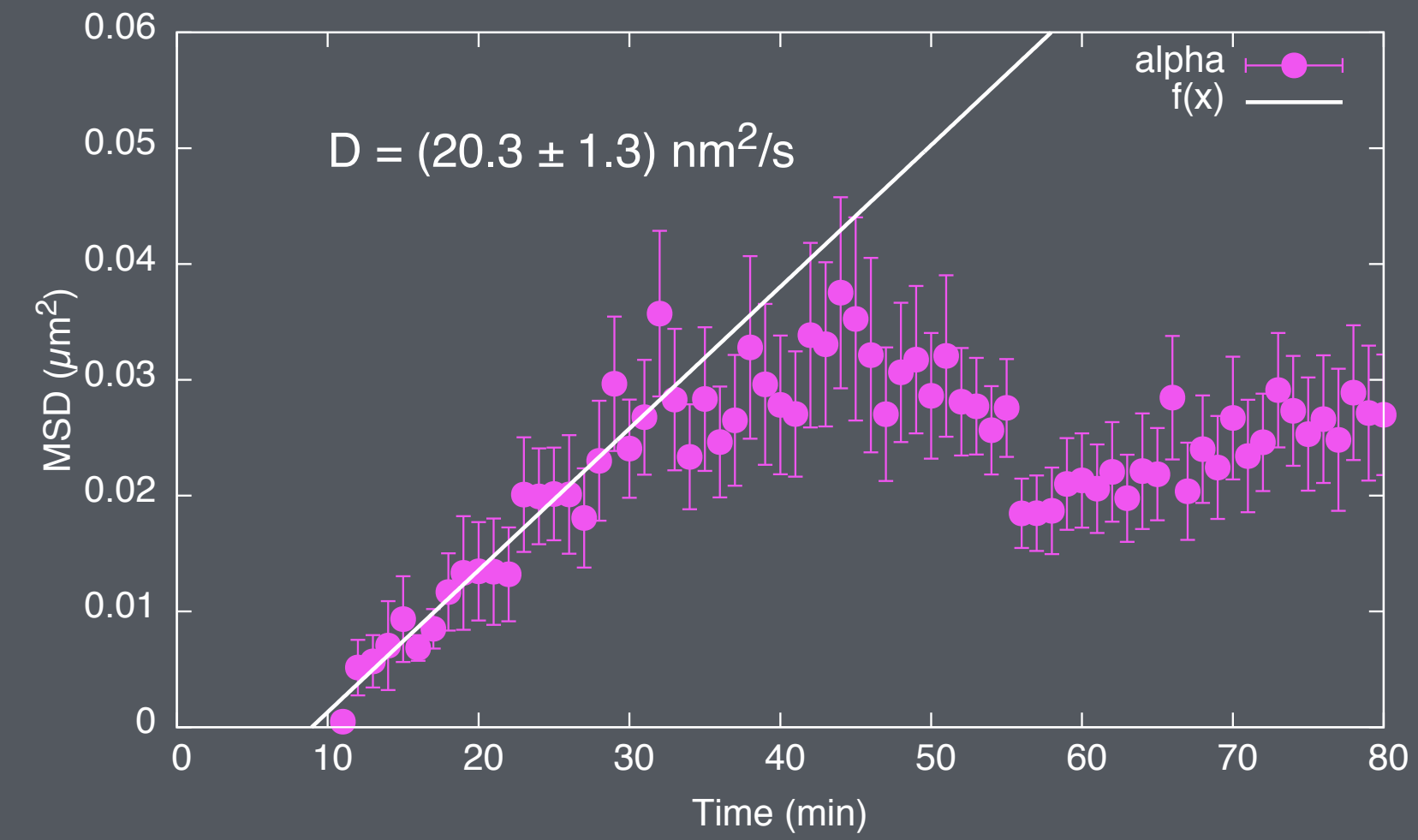
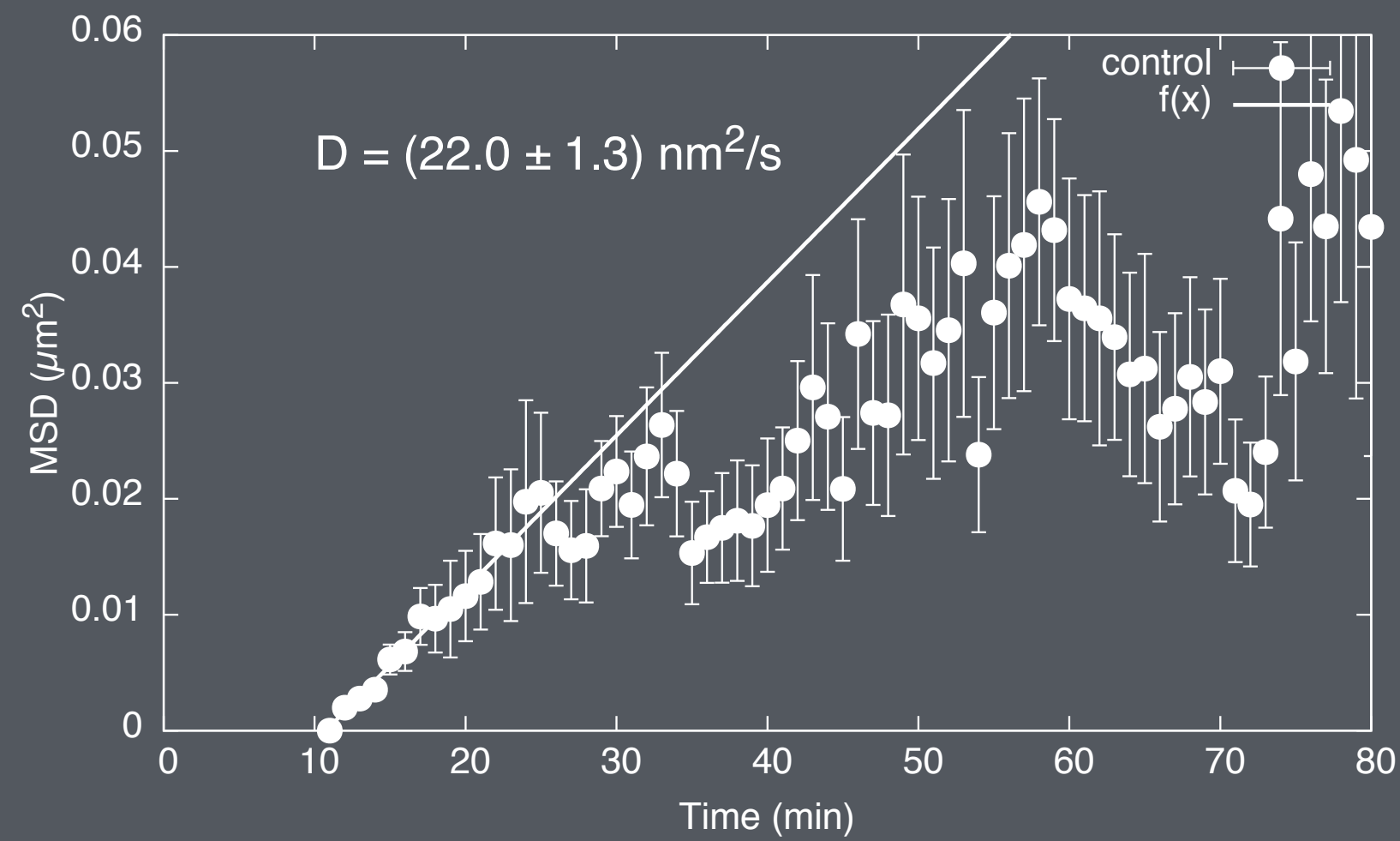
Slow repair

control: 1.9 ± 1.0
 X-rays: 5.2 ± 1.8
 alpha: 10.9 ± 2.1
 mixed: 6.7 ± 1.9

Live imaging

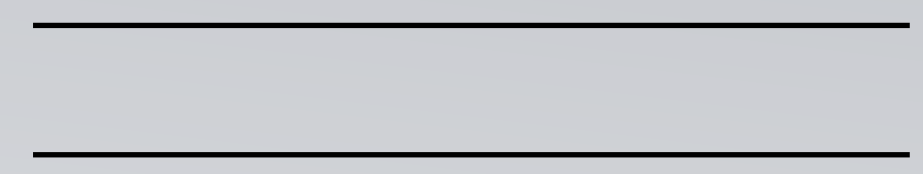
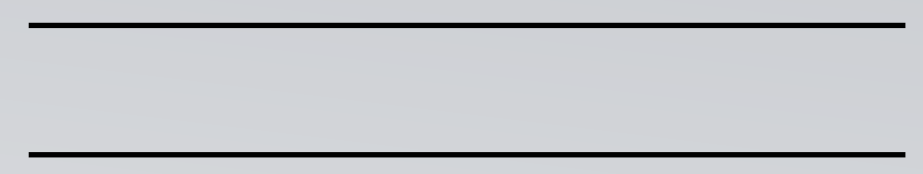


Mean square displacement

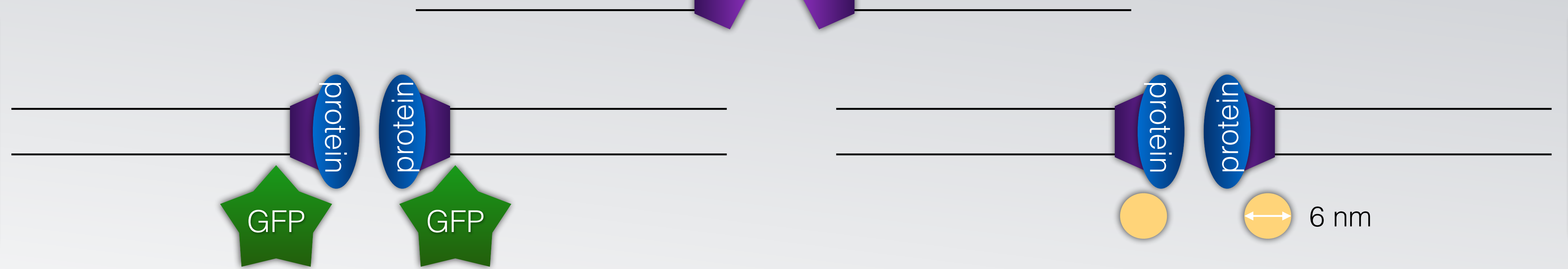
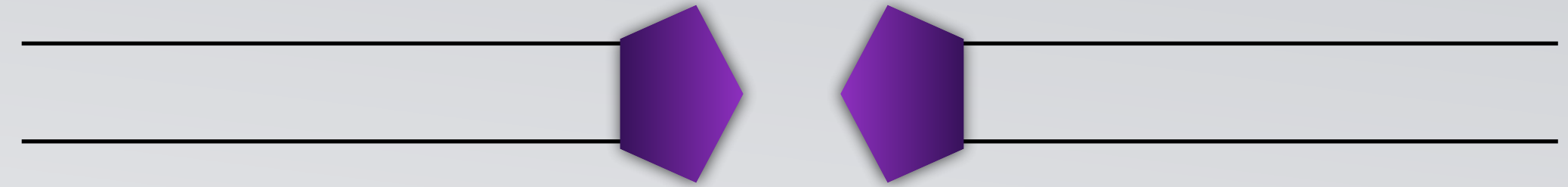


How to visualize DSB?

double strand break



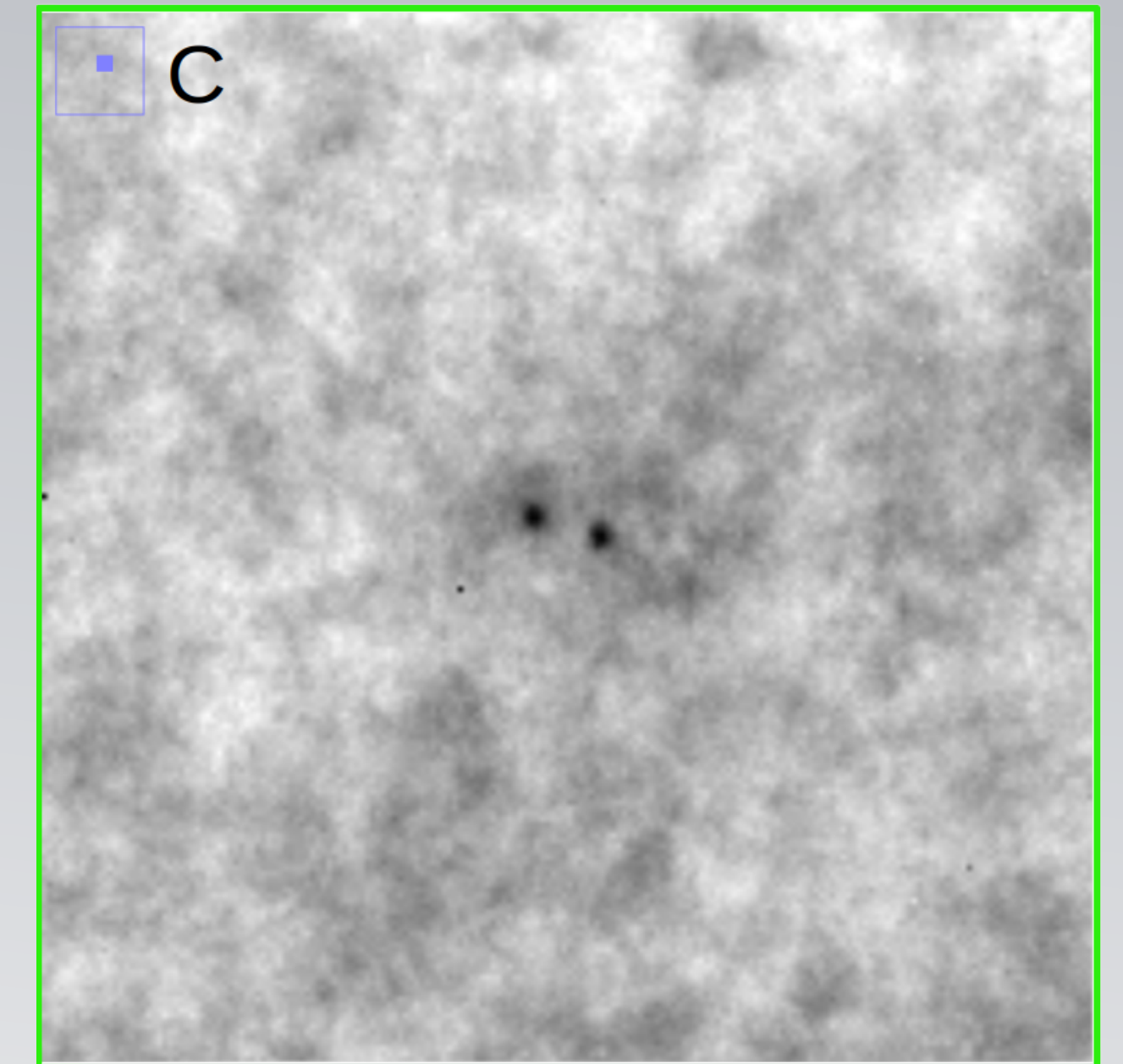
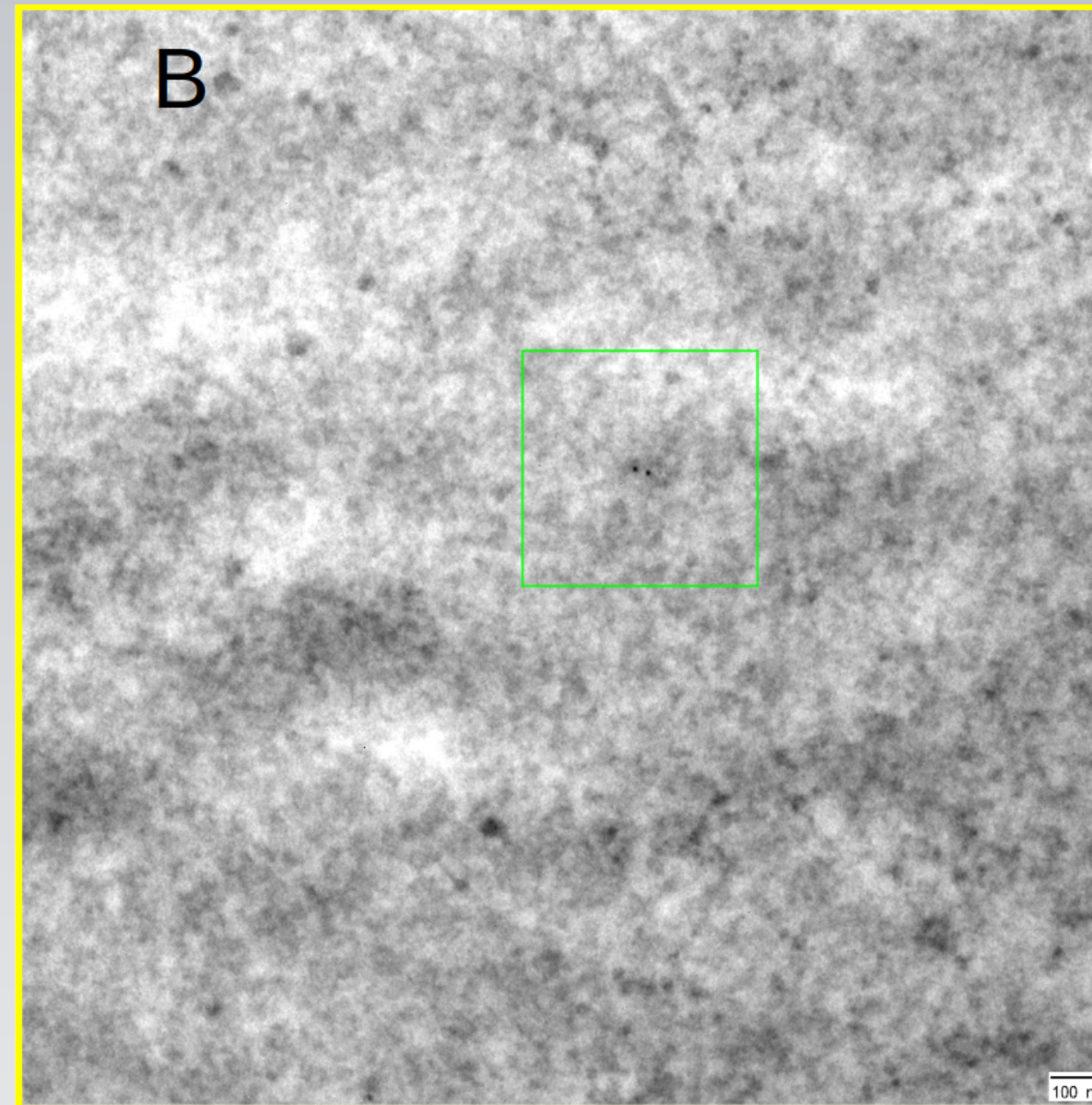
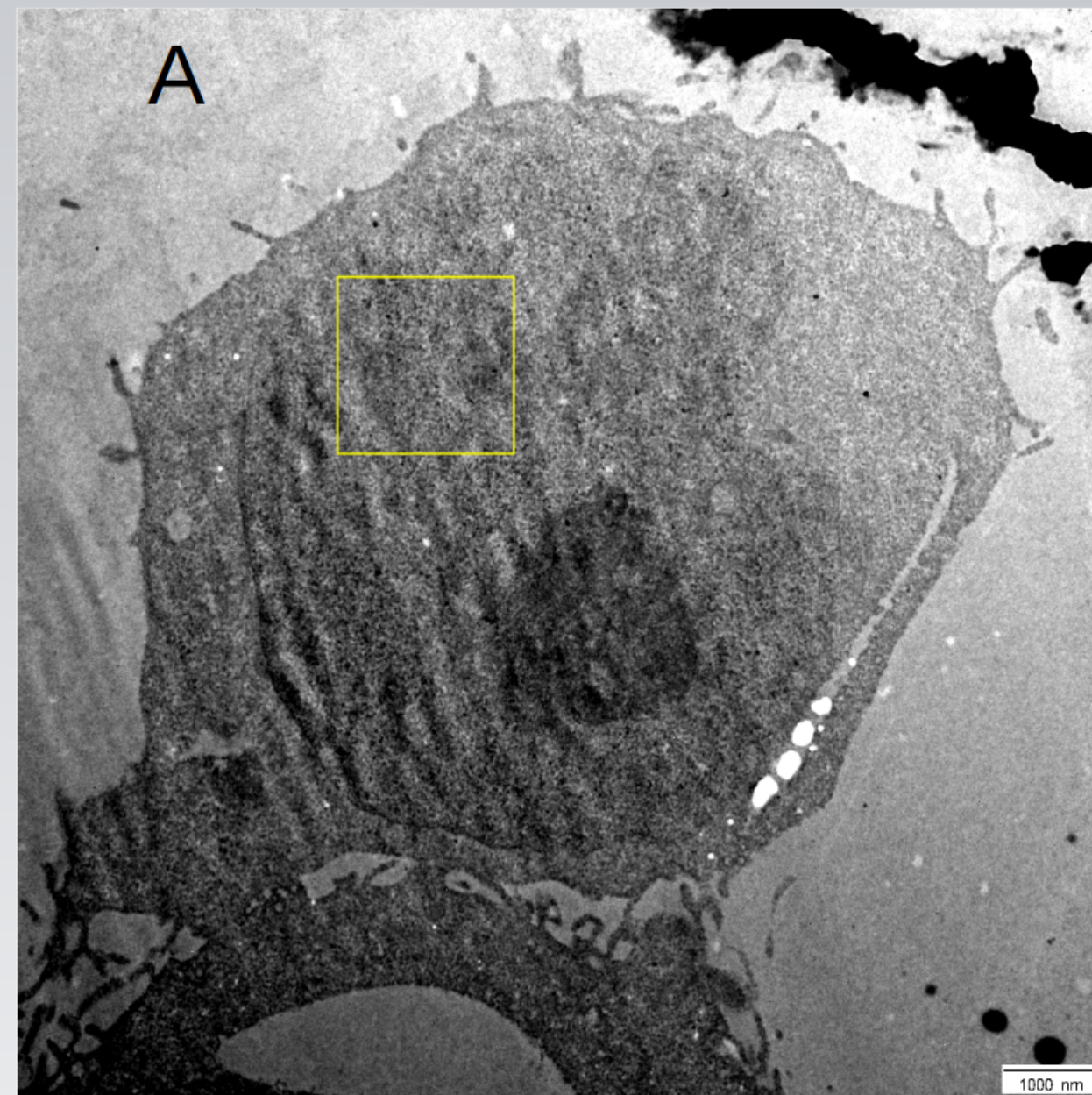
}DNA strand



focus analysis

TEM analysis

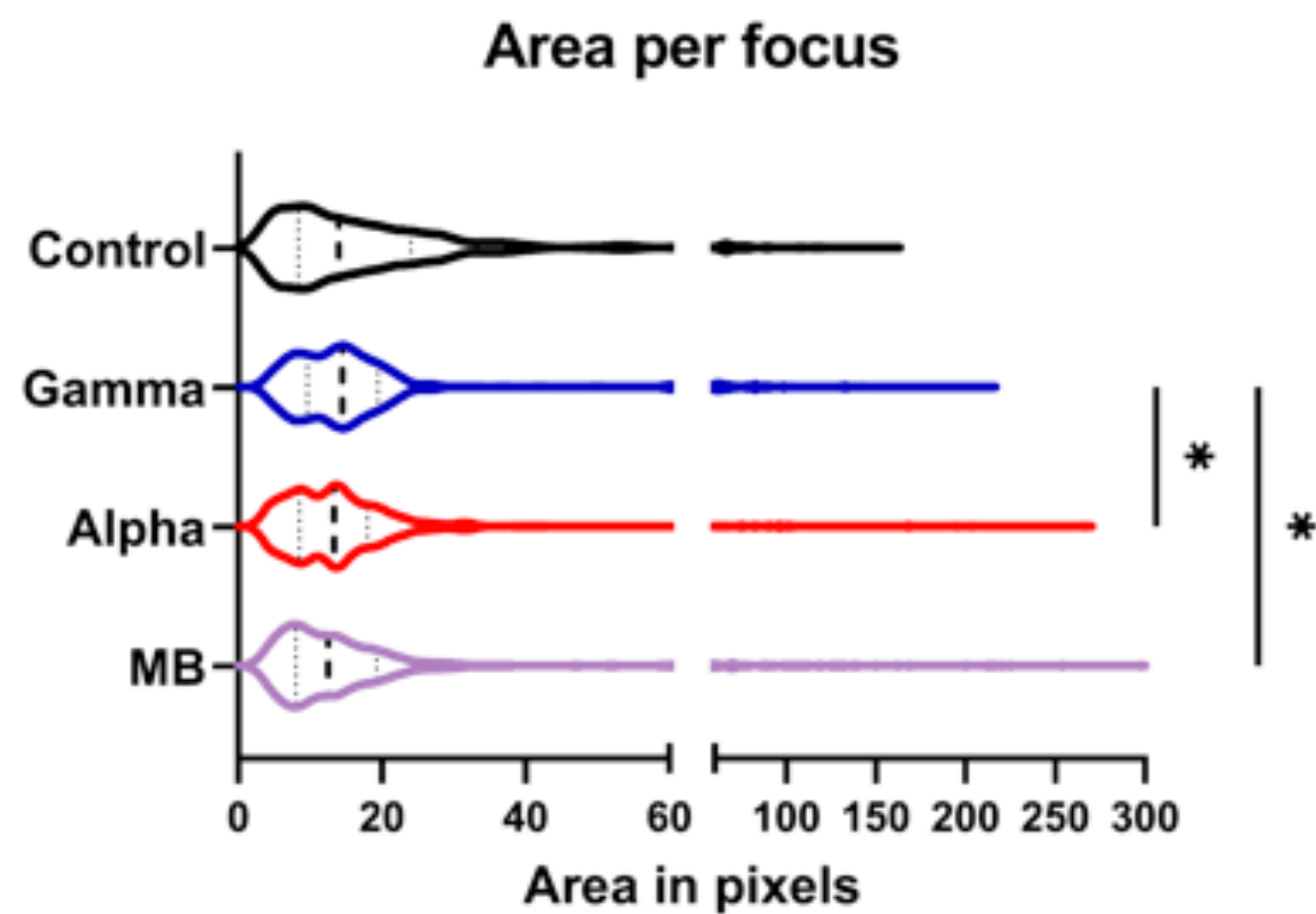
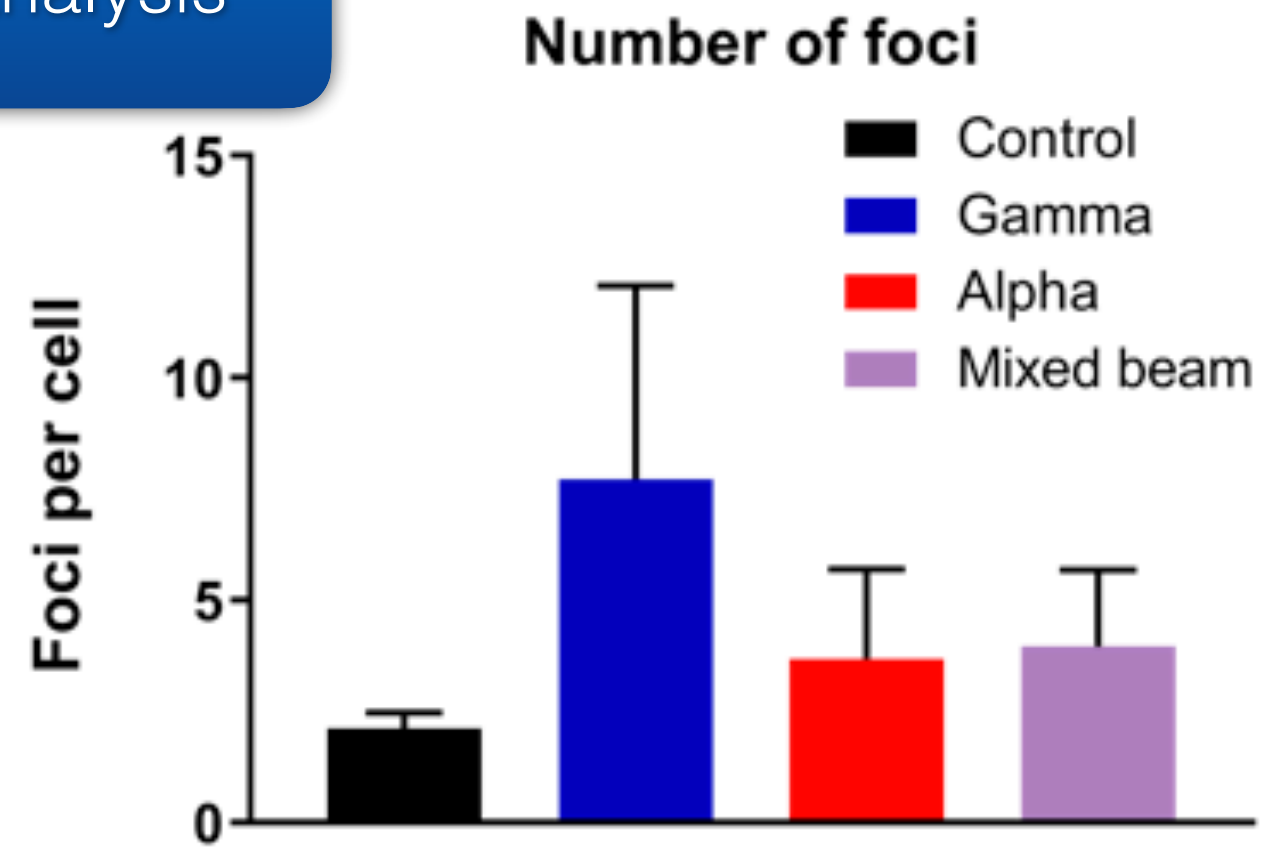
TEM analysis



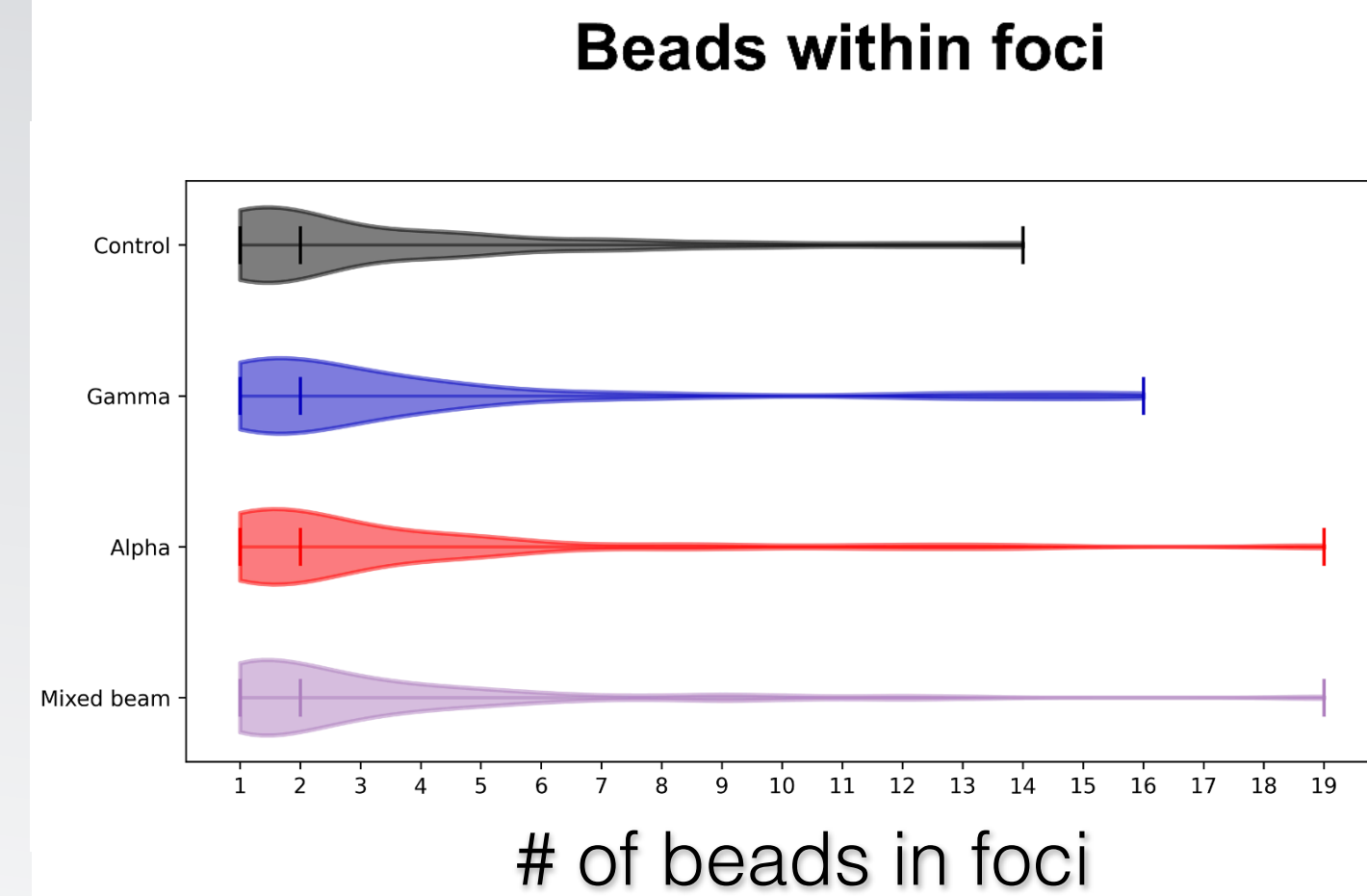
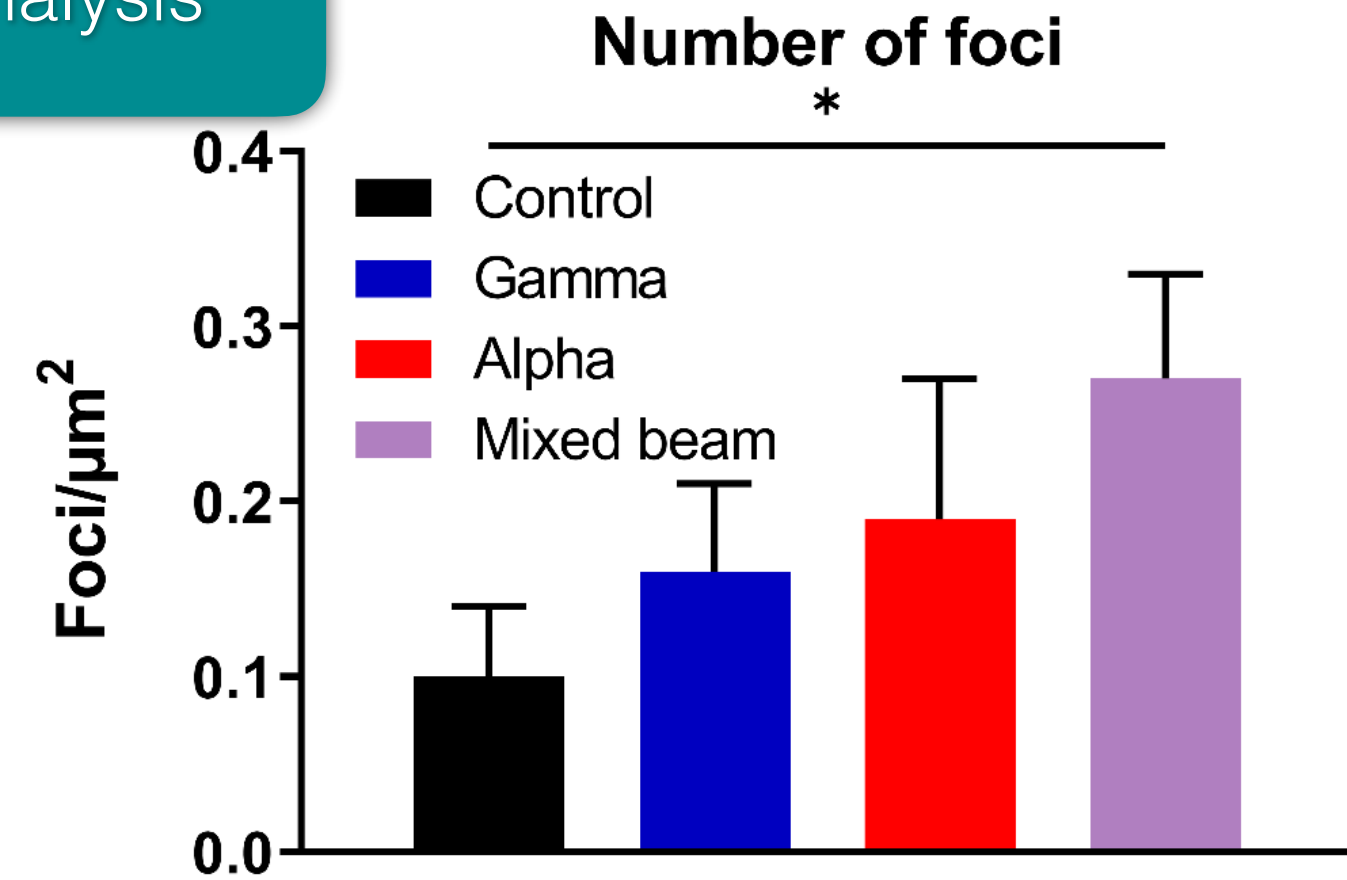
- 80 images were taken per radiation quality;
- Every picture was treated as a random representation of all investigated areas;
- Images were normalised to obtain a mean value of pixel darkness equal to 0 and a standard deviation equal to 1;

Focus number and size

γ H2AX analysis

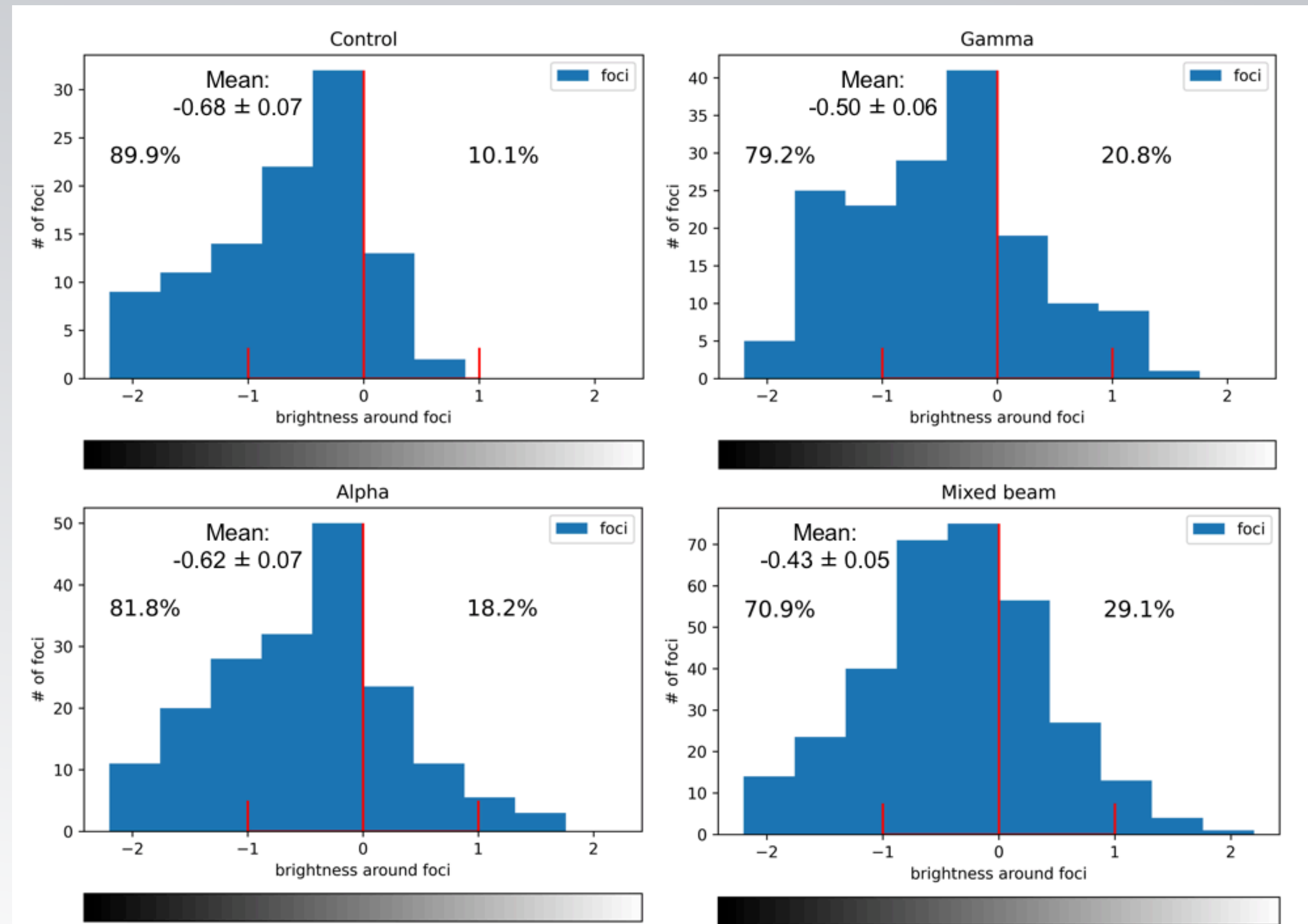
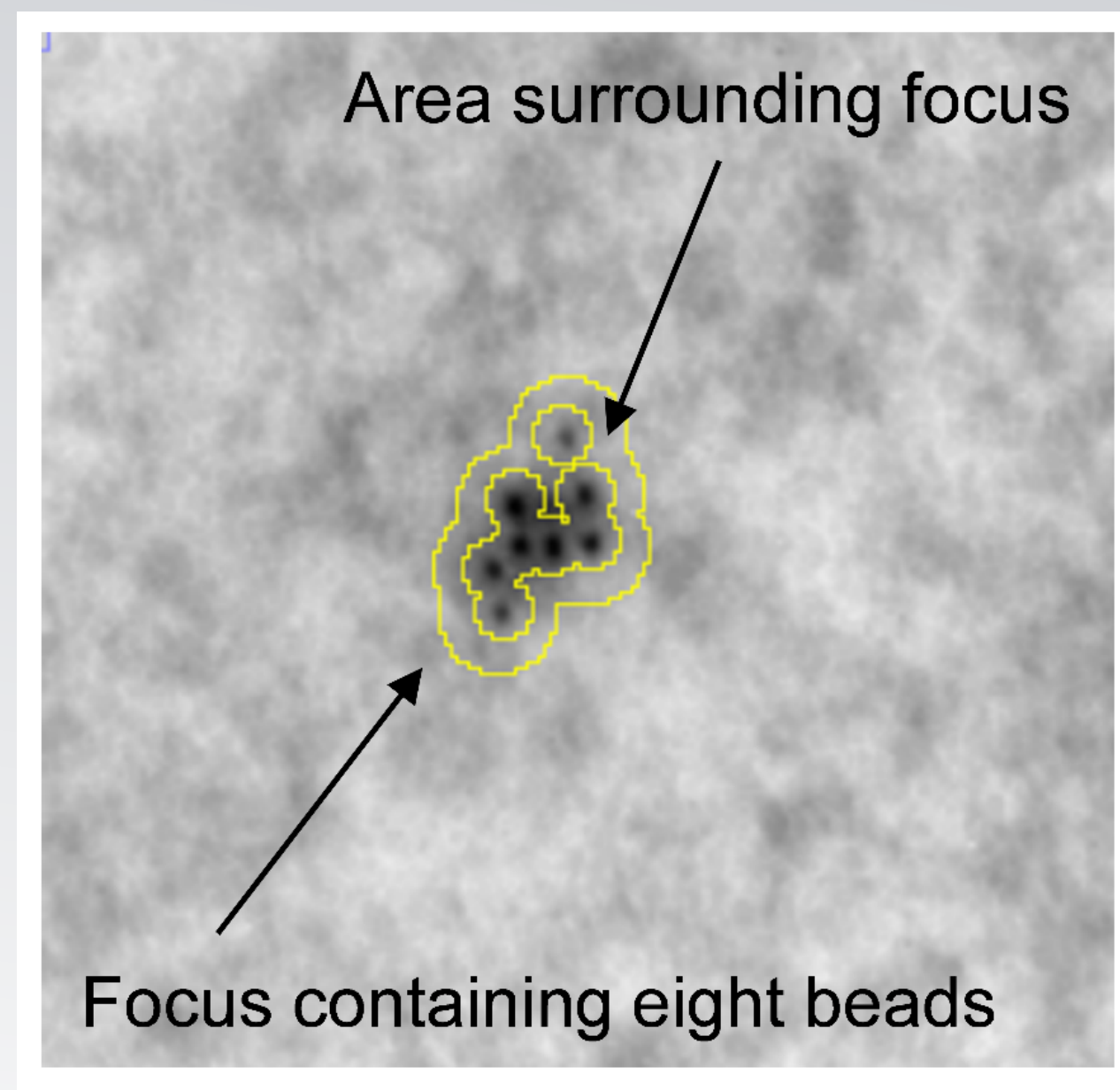


TEM analysis



Focus localisation within chromatin

For each area surrounding a bead or focus (between the outer and inner ring), the average pixel intensity (brightness) was computed



Conclusions

Damage response machinery

- additional damage induced by the second agent may not be repaired properly or in the usual time frame;
- the pool of freely available repair proteins is depleted by high LET IR;
- initial elimination of foci induced by mixed beams proceeded more slowly than expected;

Synergistic effect

- **high and low LET radiations interact to produce more DNA damage than expected from an additive action;**
- the mixed beam effect should be considered when transferring cancer risk and radiation protection.



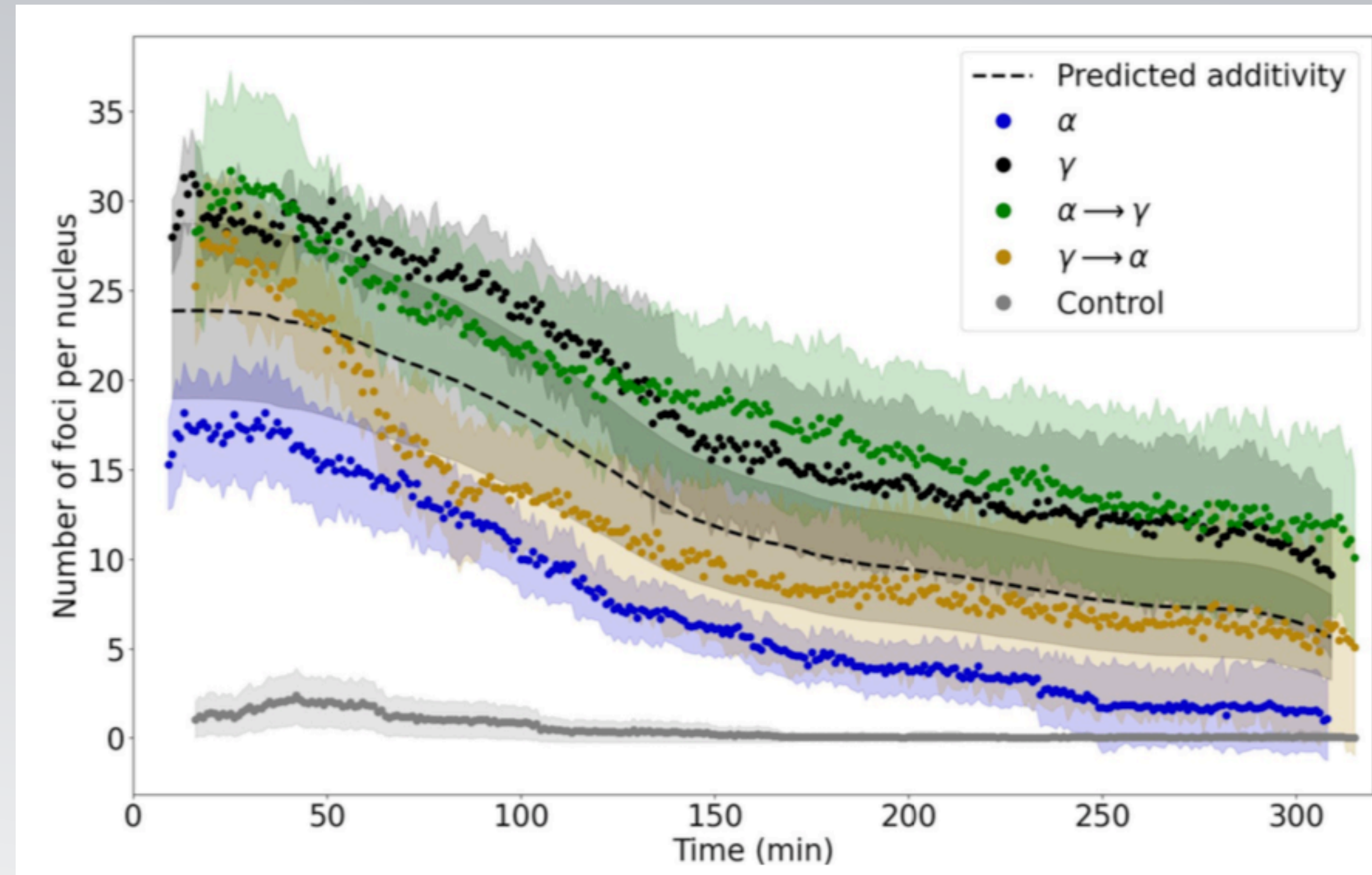
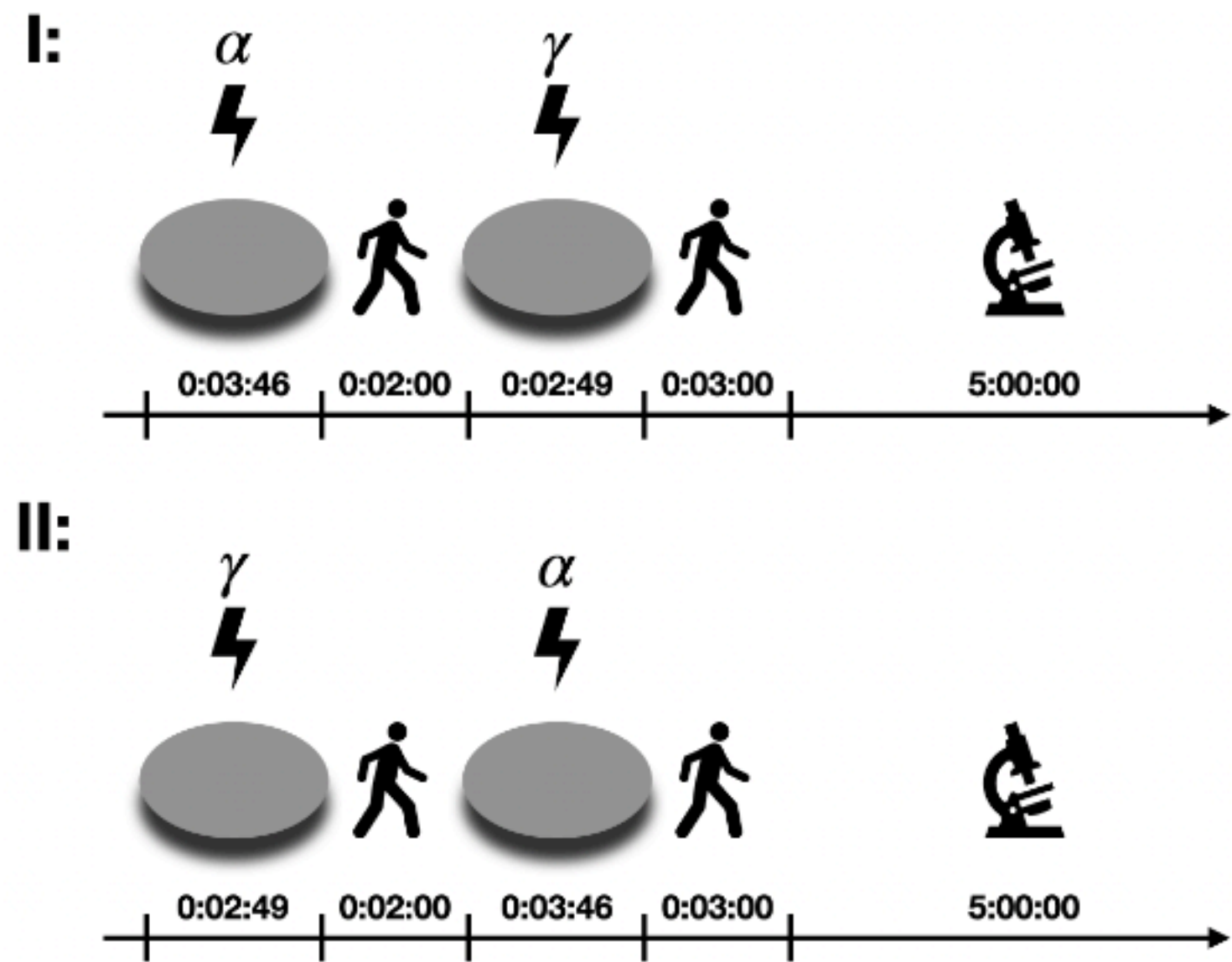
Department of Molecular Biosciences, The Wenner-Gren Institute

Stockholm
University



Andrzej Wójcik
Lei Cheng
Lovisa Lundholm
Alice Sollazzo

Does the order of irradiation matter: NBS1 focus dynamics



Adrianna TARTAS

Medical physics

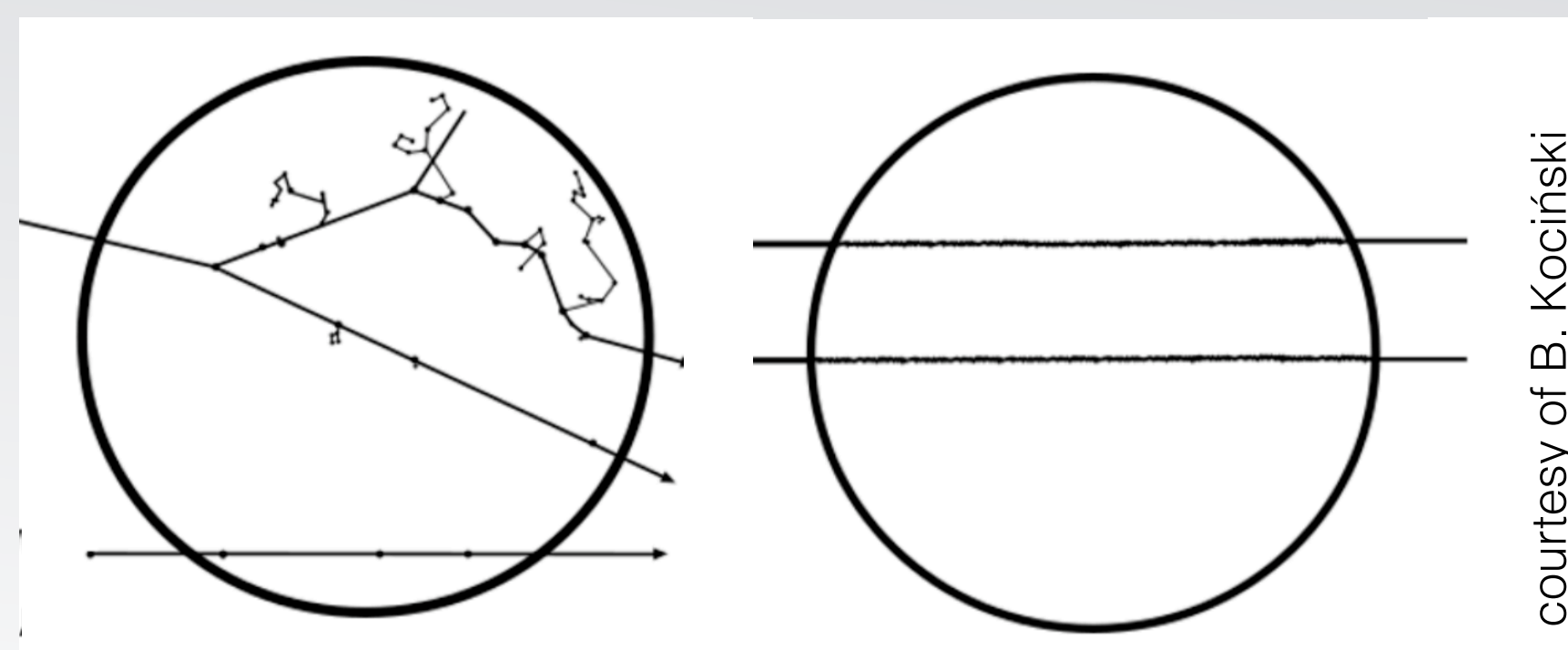
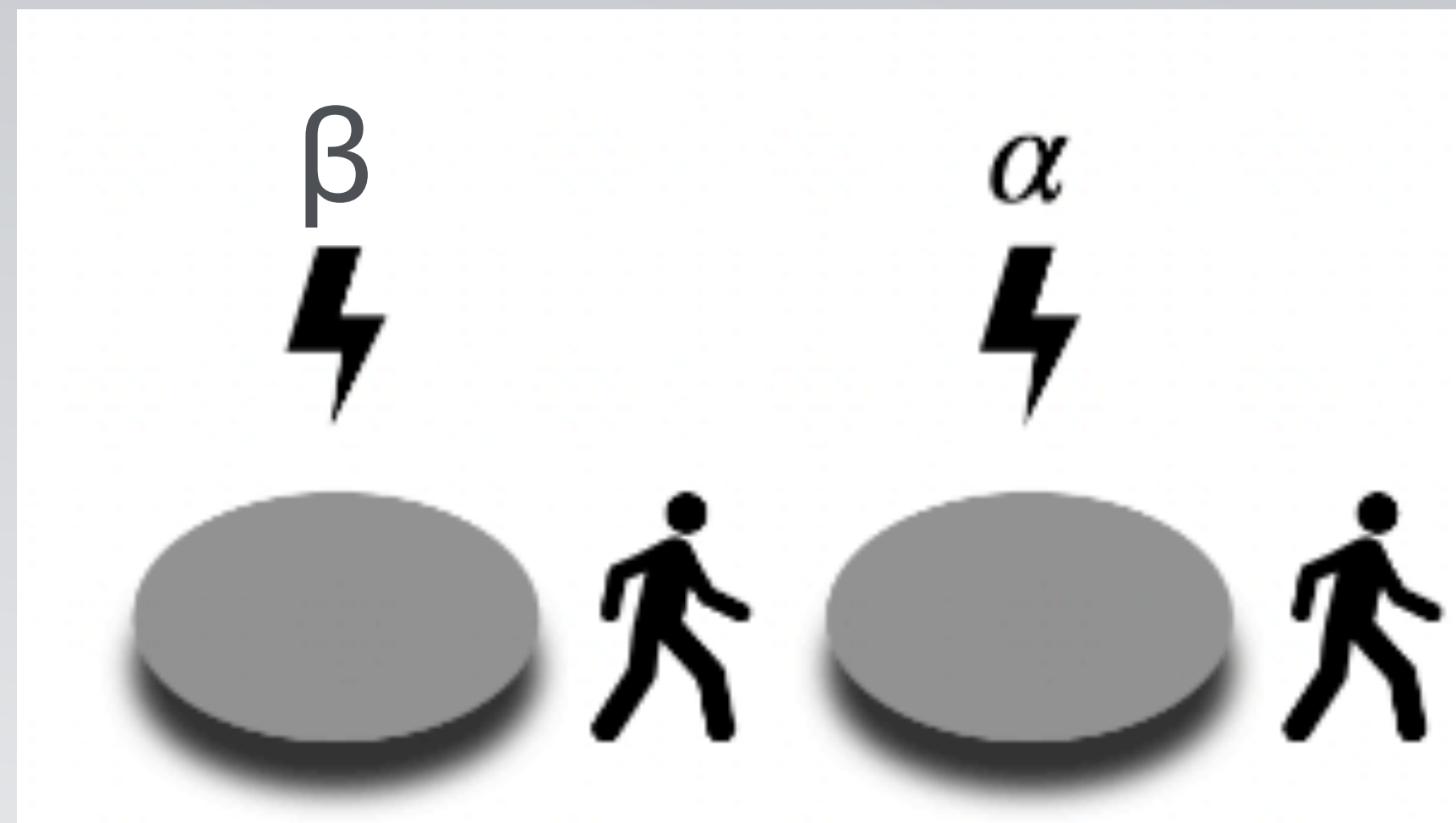
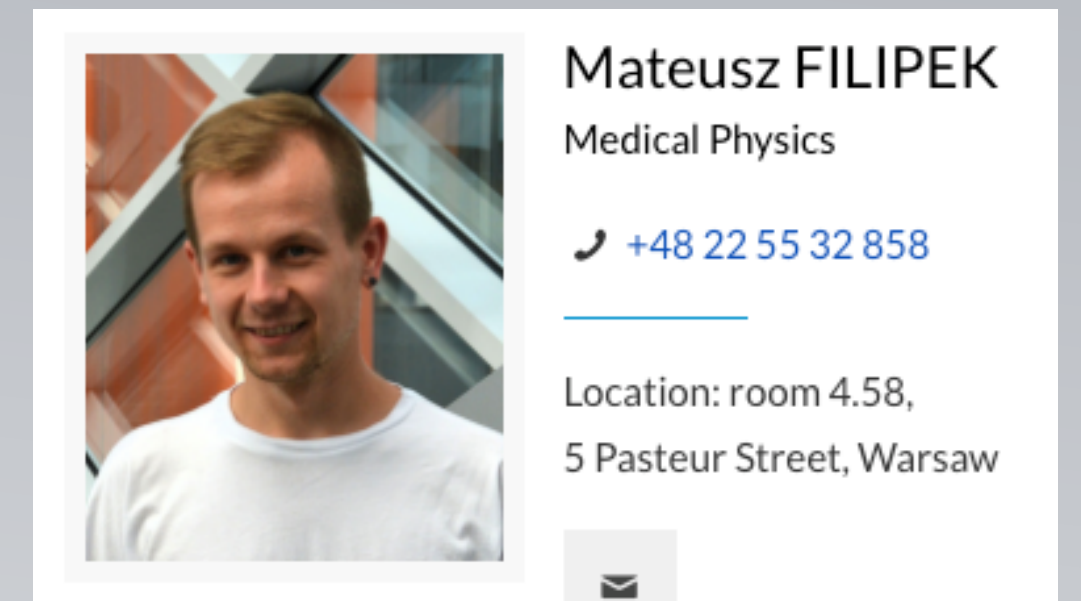
+48 22 55 32 858

Location: room 4.58,
5 Pasteur Street, Warsaw



Tartas A., Lundholm L., Scherthan H., Wojcik A. i Brzozowska B. (2023). The order of sequential exposure of U2OS cells to gamma and alpha radiation influences the formation and decay dynamics of NBS1 foci, PLoS ONE 18(6): e0286902 (<https://doi.org/10.1371/journal.pone.0286902>)

Do cells adapt to radiation?



250 mGy

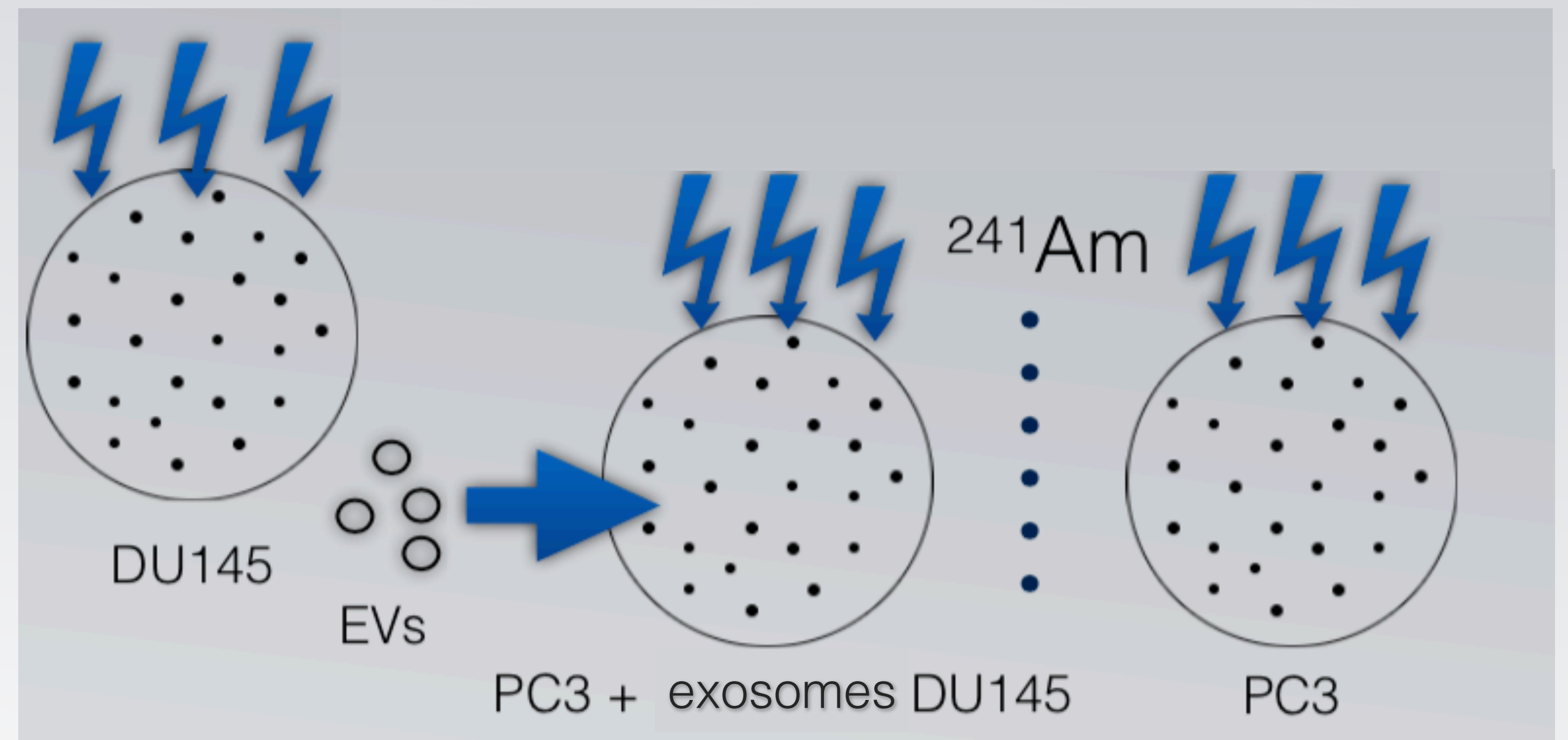
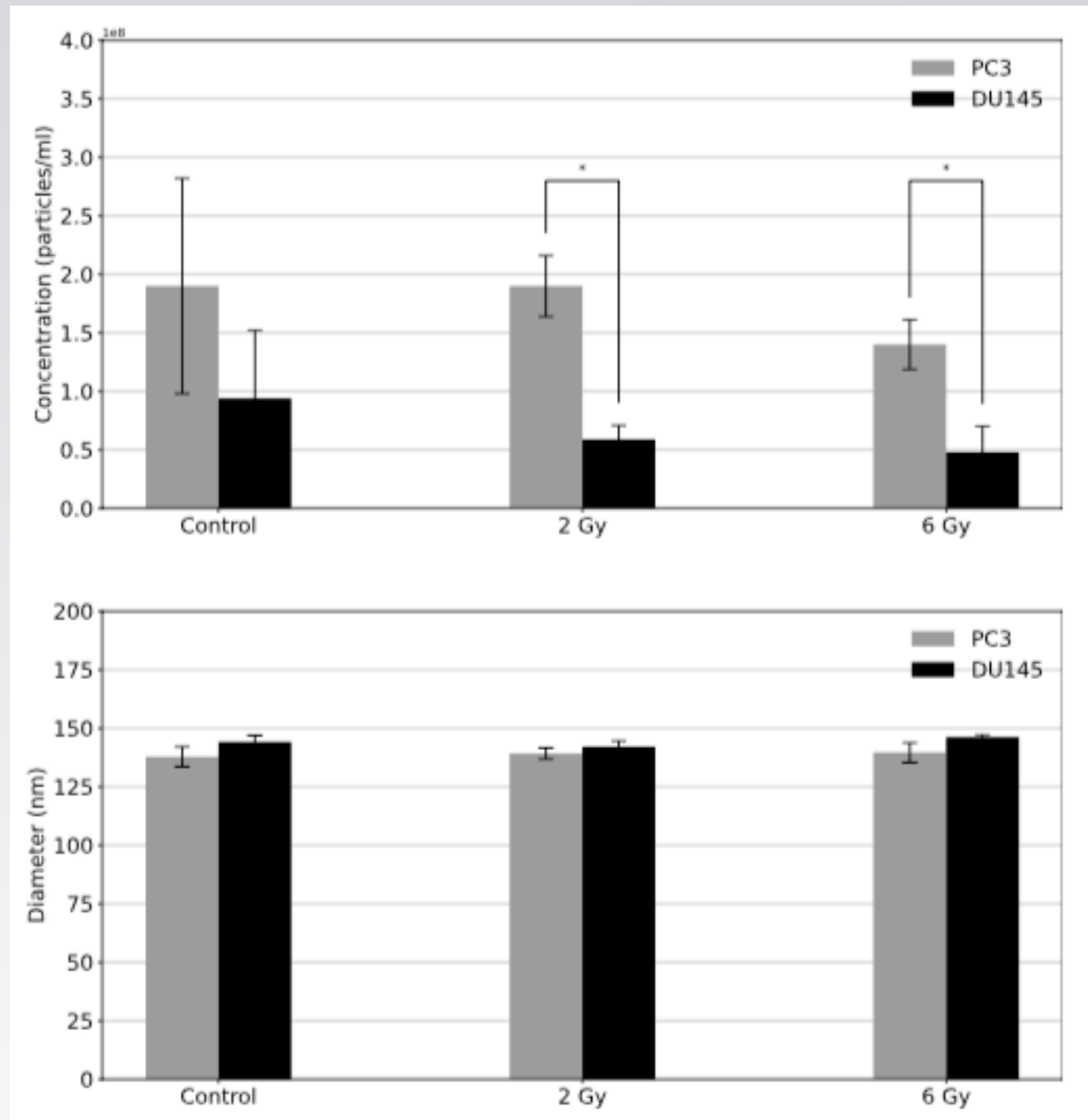
2 Gy

courtesy of B. Kociński

CELL SURVIVAL

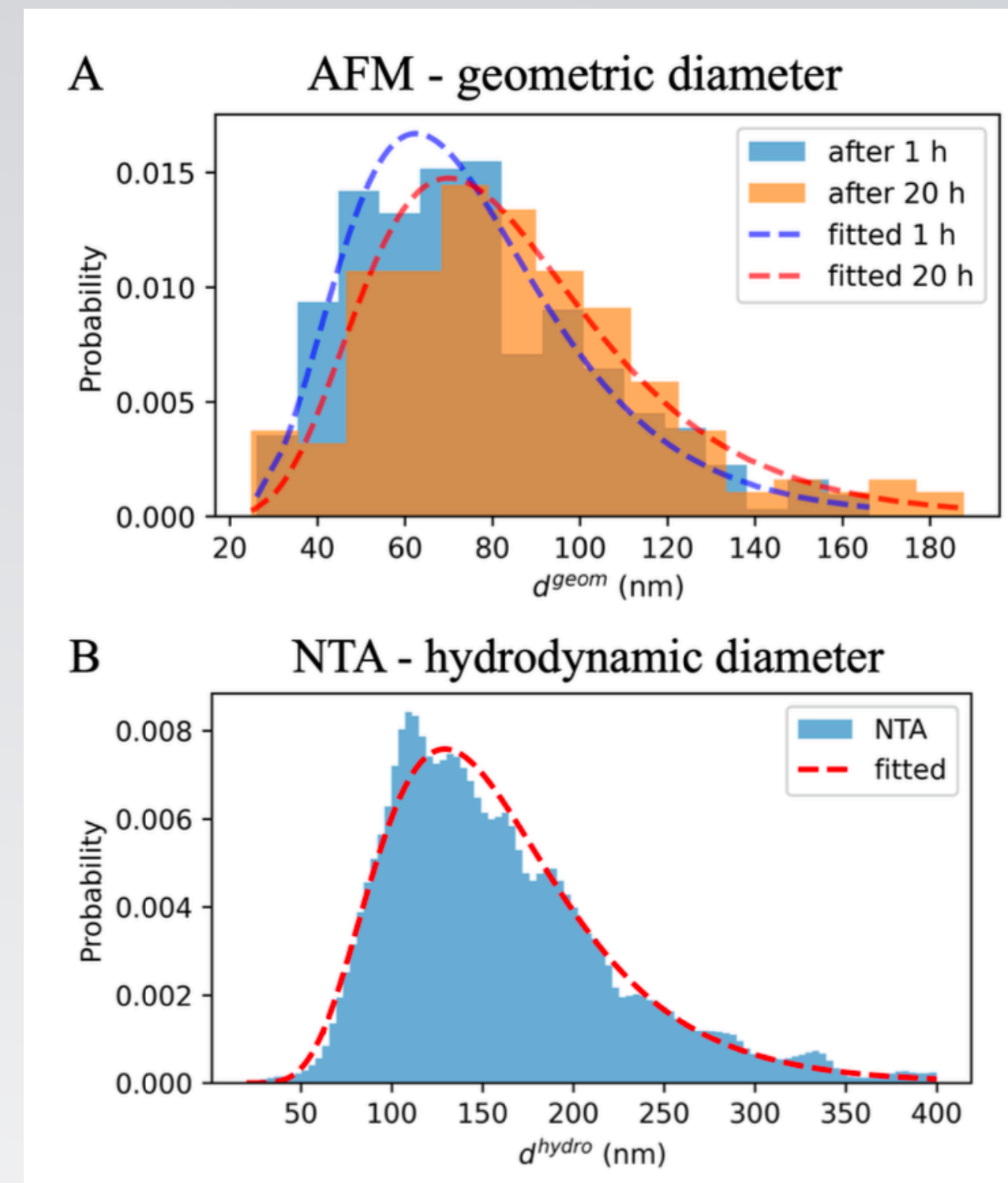
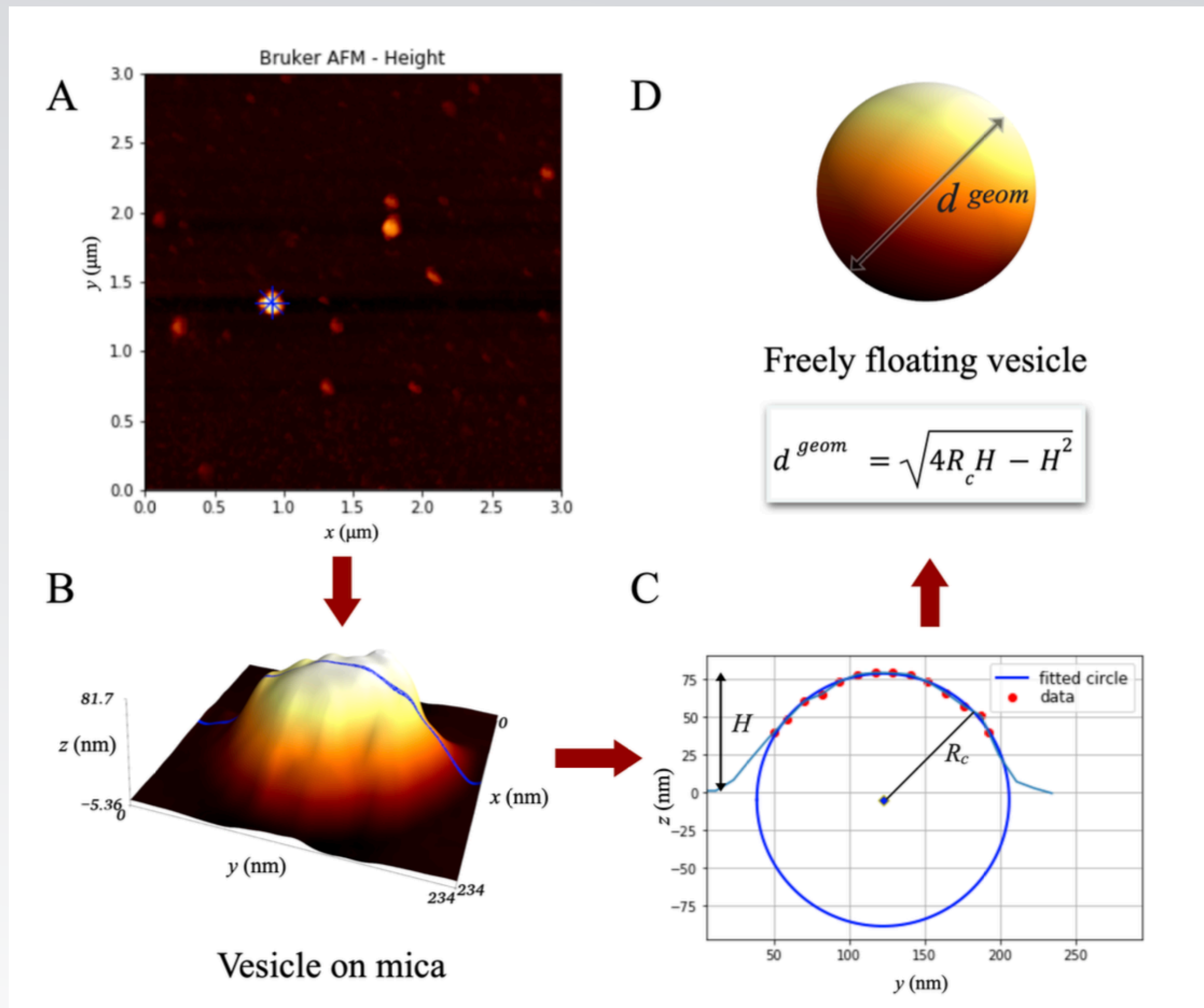
REPAIR FOCUS ANALYSIS

Do exosomes make cells more radiosensitive?




Pszczółkowska B., Olejarz W., Filipek M., Tartas A., Kubiak-Tomaszewska G., Żoźnierzak A., Życieńska K., Ginter J., Lorenc T. i Brzozowska B. (2022). Exosome secretion and cellular response of DU145 and PC3 after exposure to alpha radiation. Radiation and environmental biophysics, 1-12

Can we distinguish between exosomes from healthy and cancer cells?



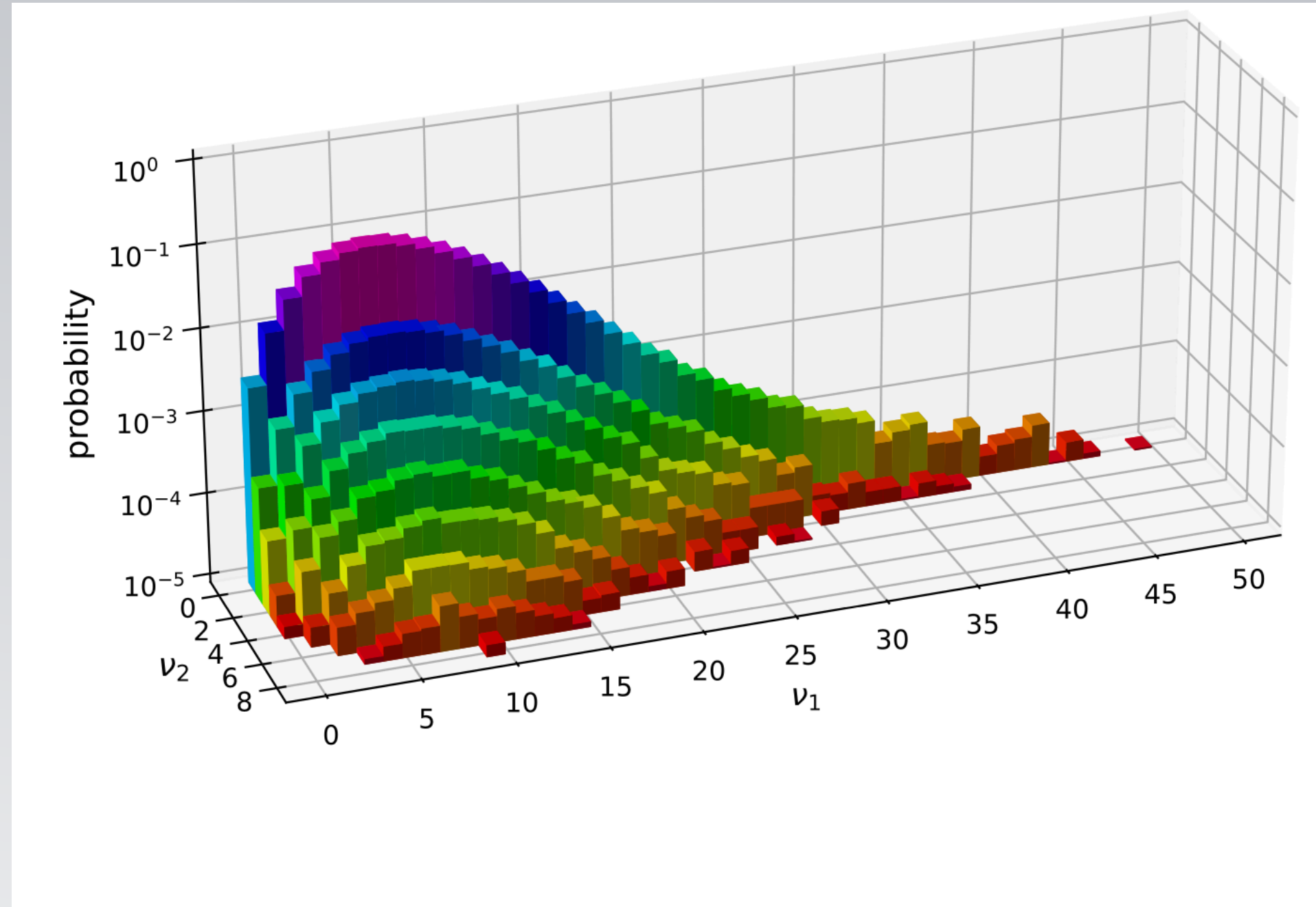
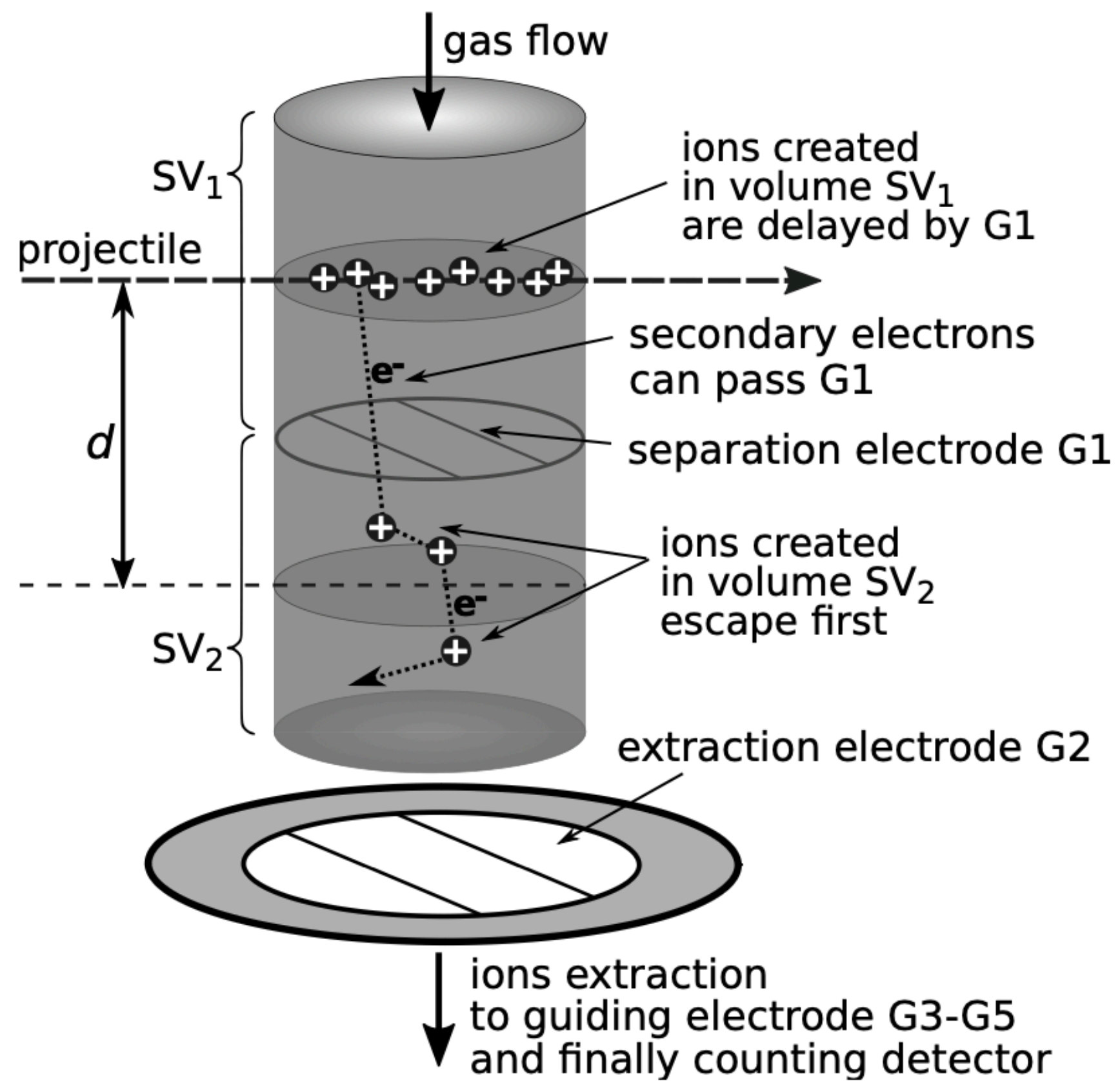

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 Supervision

Życieńska, K., Pszczołkowska, B., Brzozowska, B., Kamiński, M., Lorenc, T., Olejarczyk, W., Sęk, S. & Ginter, J. (2022). Brownian Motion Influence on AFM Exosomes' Size Measurements. International Journal of Molecular Sciences, 23(17), 10074.

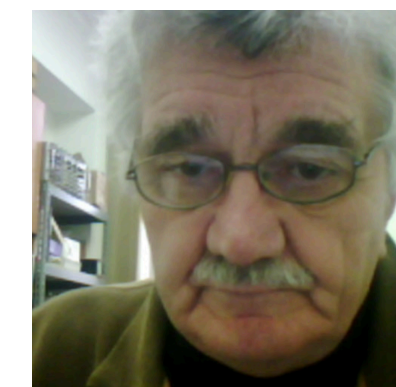
Can we measure spatial dependencies of ionisation events?



Marcin
PIETRZAK
Medical physics

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5 Pasteur Street, Warsaw



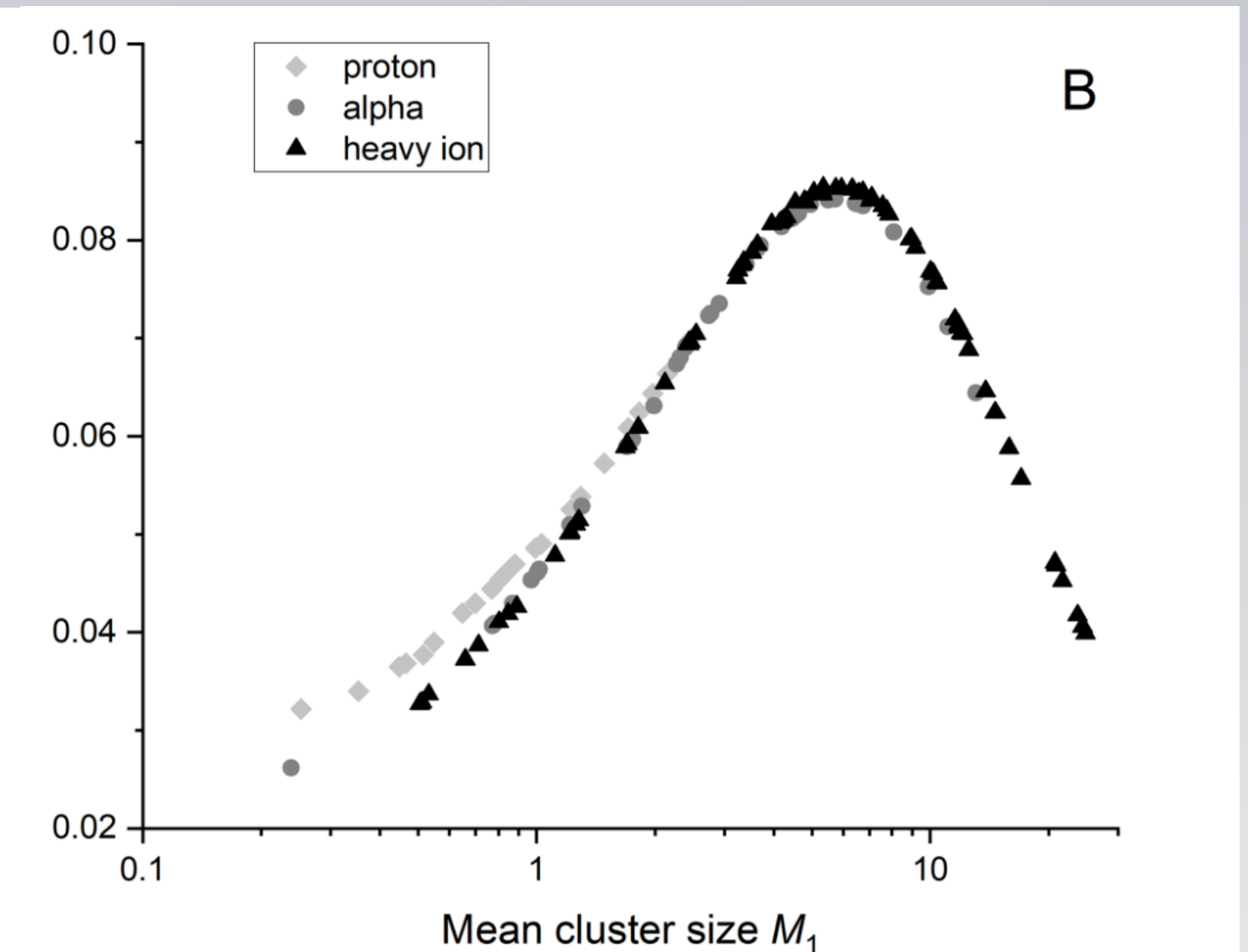
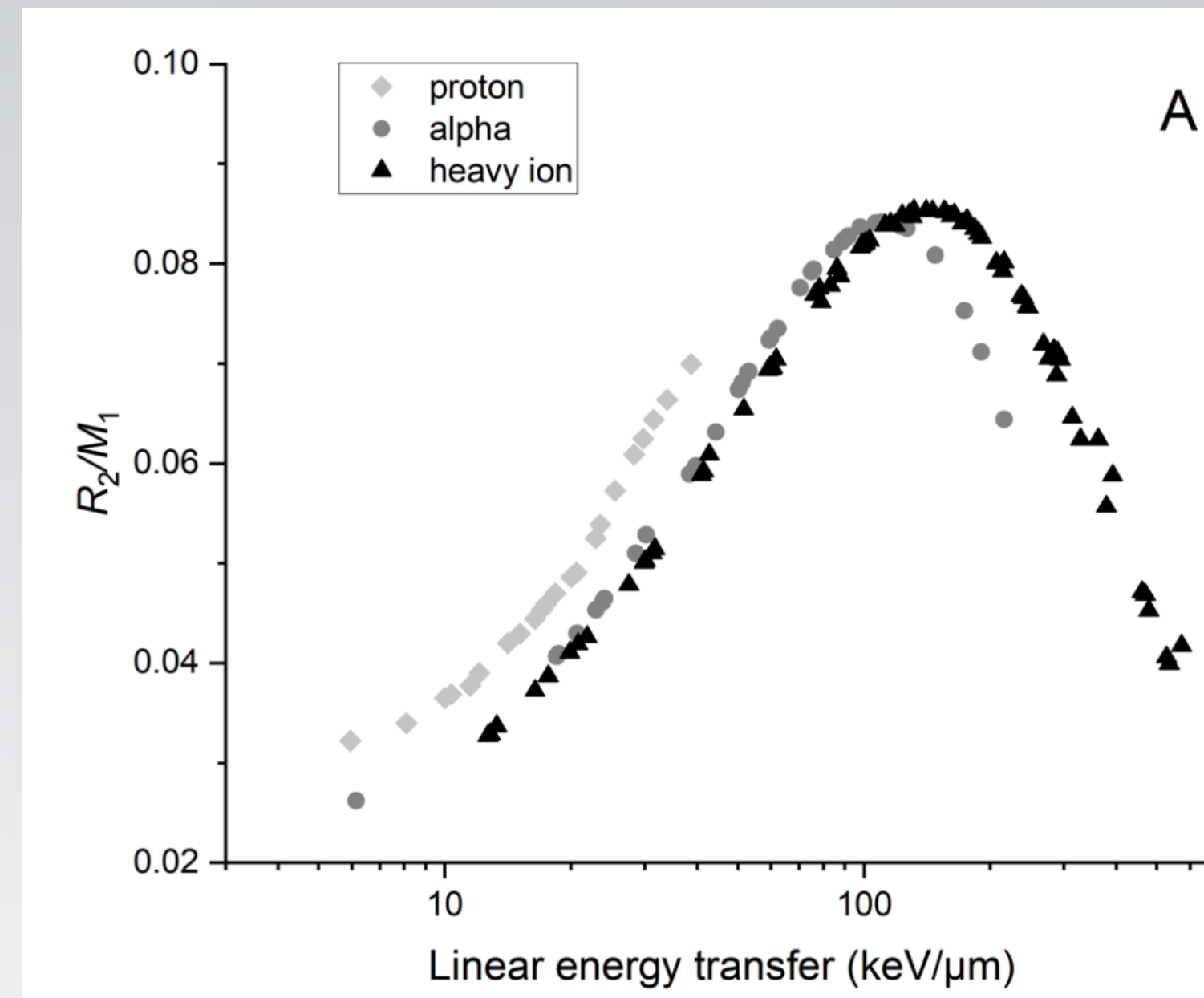
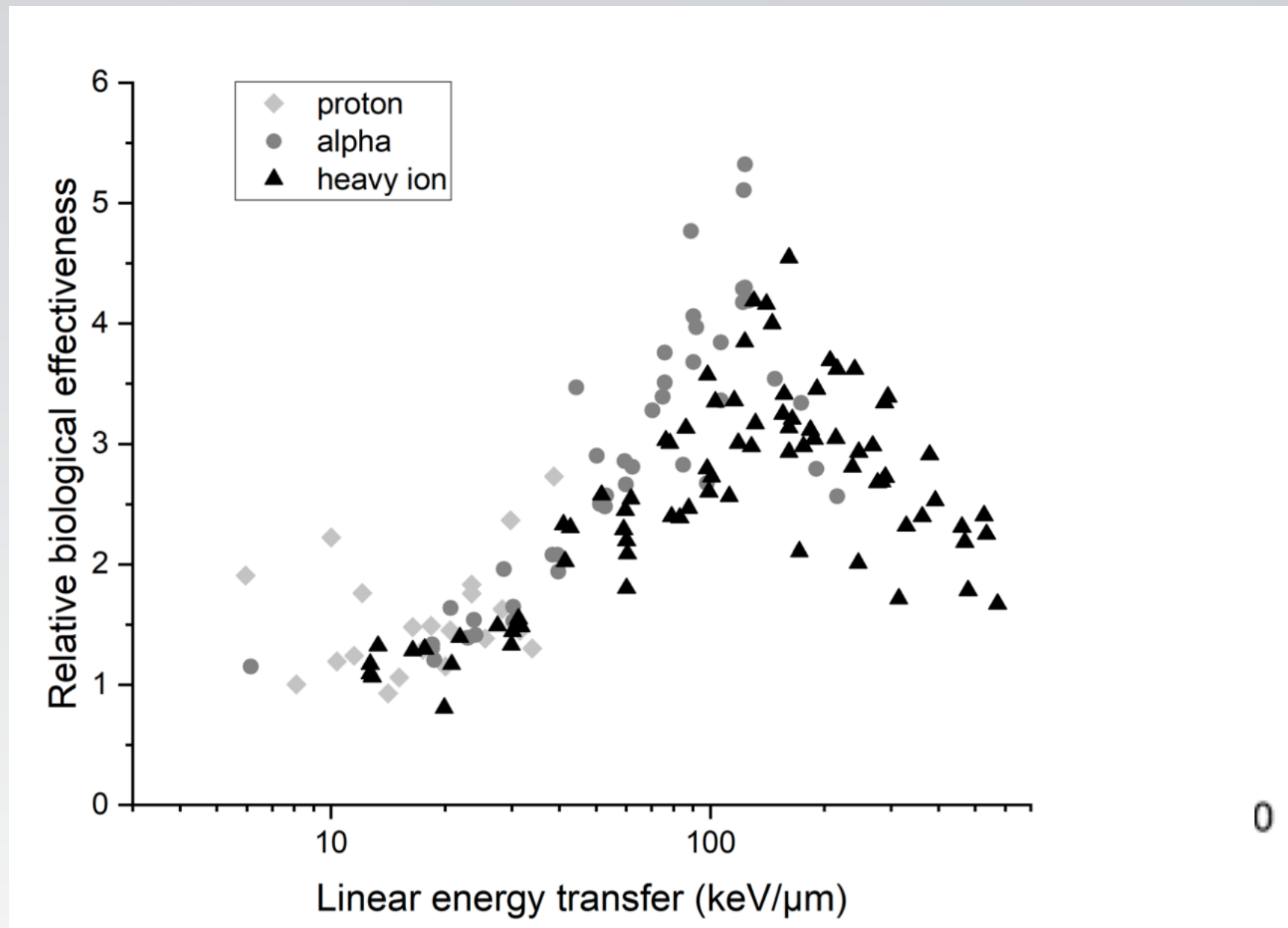
Zygmunt
SZEFLIŃSKI
Heavy Ion Laboratory

Supervision

Can nanodosimetric quantities be used to describe DNA damage?



Monika
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Location: room 4.58,
5 Pasteur Street, Warsaw



M. Mietelska, M. Pietrzak, A. Bancer, A. Ruciński, Z. Szepliński, B. Brzozowska, Ionisation detail parameters for DNA damage evaluation in charged particle radiotherapy: simulation study based on cell survival database, under preparation



*You can design and create, and build
the most wonderful place in the
world. But it takes people to make
the dream a reality.*

 Walt Disney

Thank you for your attention!
beata.brzozowska@fuw.edu.pl